

JuLI™ *Br*

Live cell movie analyzer



JuLI™ Br, Live cell movie analyzer

Instructions for using JuLI™ Br, live cell movie analyzer described below.
For detailed instructions, refer to the manual supplied with JuLI™ Br or download the manual from www.nanoentek.com

1. Starting JuLI™ Br

- ▶ Turn on the **Power button**.
Place the sample on the stage.

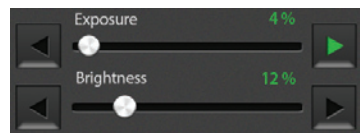


2. Focusing

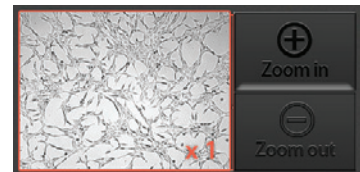
- ▶ Adjust the focus for each scope unit if 2nd unit is also connected to station unit. Press **Channel tab** to select each scope unit.



- ▶ Adjust the illumination intensity using the **Exposure** and **Brightness bar** of Focusing menu.



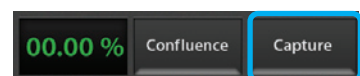
- ▶ Set the viewing region of image by pressing the **Zoom in/out button** and **dragging preview window**.



- ▶ Adjust focus of the image using the **Focus knob** or **Focus interface**.

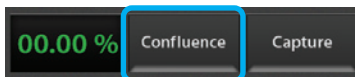


- ▶ Press the **Capture** and **Save button** to acquire the image if it is necessary.



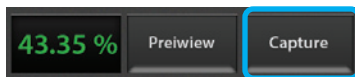
3. Getting confluence value

- ▶ Press **Confluence** button of Focusing menu.



If the value of confluence should be more accurate, try again after adjusting **parameters** of Settings menu.

- ▶ Press the **Capture** and **Save** button to acquire the image with confluence circle if it is necessary.



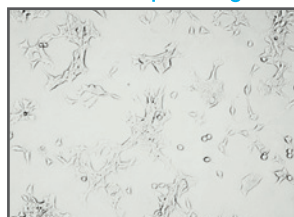
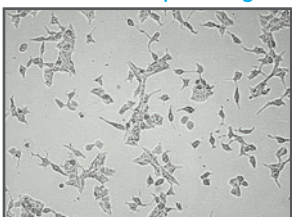
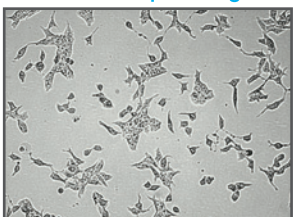
The reference images for correct focus and confluence

Du-145 (Parameter A)

Good sample image

Normal sample image

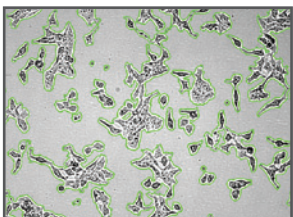
Poor sample image



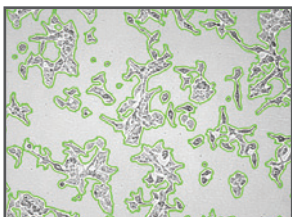
Reference: Exposure 4 / Brightness 1

Reference: Exposure 6 / Brightness 3

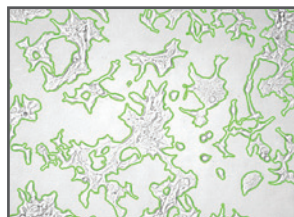
Reference: Exposure 6 / Brightness 5



Confluence 35.11%



Confluence 36.22%



Confluence 43.03%

Parameter	Cell line	Recommendation
	A	HepG2, GH3, Hep3B, A549, MCF7, SH-SY5Y, SCN2.2, NIH-3T3, F9, HeLa, Du-145
	B	LNCaP
	C	U-2 OS, ADSC
		Sensitivity level: 9 Background level: 2
		Sensitivity level: 10 Background level: 2
		Sensitivity level: 10 Background level: 8

4. Making time lapse imaging



Warm up scope unit for 1 hour in incubator before making a movie.
If not, it might be cause of uncertain data or unclear movie.



Cell culture flask should be wetted by the culture media before making movie.

- ▶ Each scope channel can be viewed by pressing **channel tab** if 2nd scope unit is connected to station unit.

- ▶ Press **Setting button** of Monitoring menu.

- ▶ Press the **Name form** to type file name and **Save button**.

- ▶ Choose the monitoring type among **Growth rate, Wound healing** and **Movie only**.

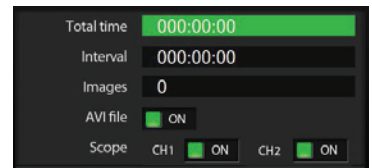
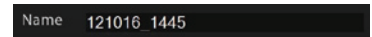
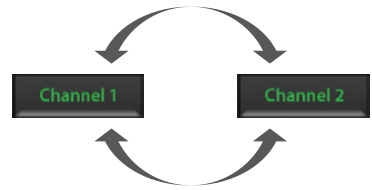
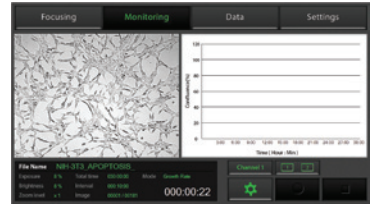
- ▶ Set up **Total time** and **Time interval**.



*Time Interval should be set up more than 1 minute for the best quality result in case of monitoring growth rate and wound healing. But In case of **dual monitoring**, more than 5 minute is recommended.*

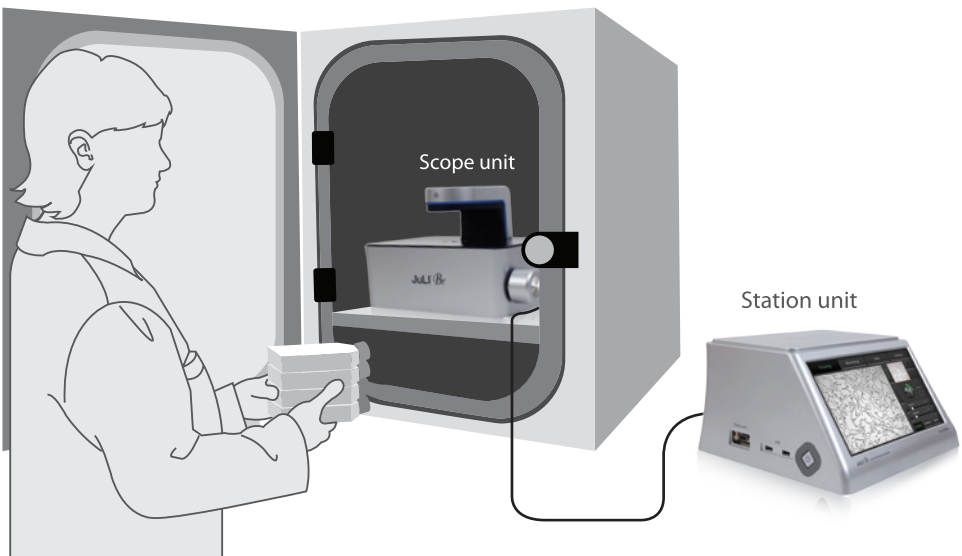
- ▶ Press the **Apply button** to save all of options.

- ▶ Press the **Rec. button** to begin recording.



○ Specification

Cat no	Device JULI-BR04 (Single set, 1 Scope & 1 Station) JULI-BRD04 (Dual set, 2 Scopes & 1 Station) JULI-BRSC (2 nd Scope) Accessory JULI-BRCM (Counting starter kit) JULI-BRTB (XY Stage)
Magnification	Objective 4 X and digital zoom (~ 400 X)
Image resolution	2560 x 1920 pixels (5M)
Exported formats	JPEG (image), AVI (movie), CSV (raw data)
Display	10.1" LCD touch screen
Light source	White LED
Dimensions & weight	Scope: 300 x 190 x 188 mm, 4 kg Station: 282 x 285 x 160 mm, 3.2 kg
Storage	320 GB Hard drive 4 GB USB drive



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Nano**EnTek**

NESMC-JUB-001E (V.1.5)

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