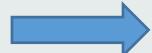


# Things you need to know about figure-making for publication



The  
Westmead  
Institute

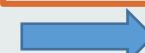
Collect data



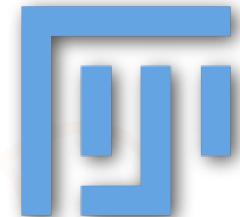
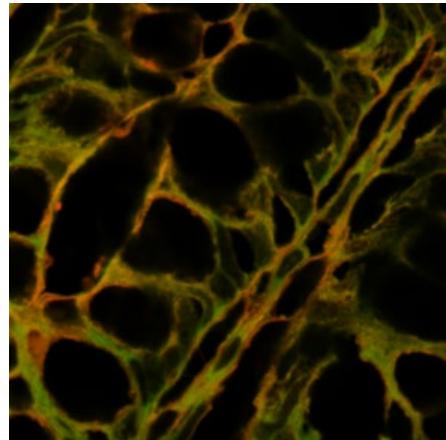
Process

Analyse

Figure



Publish



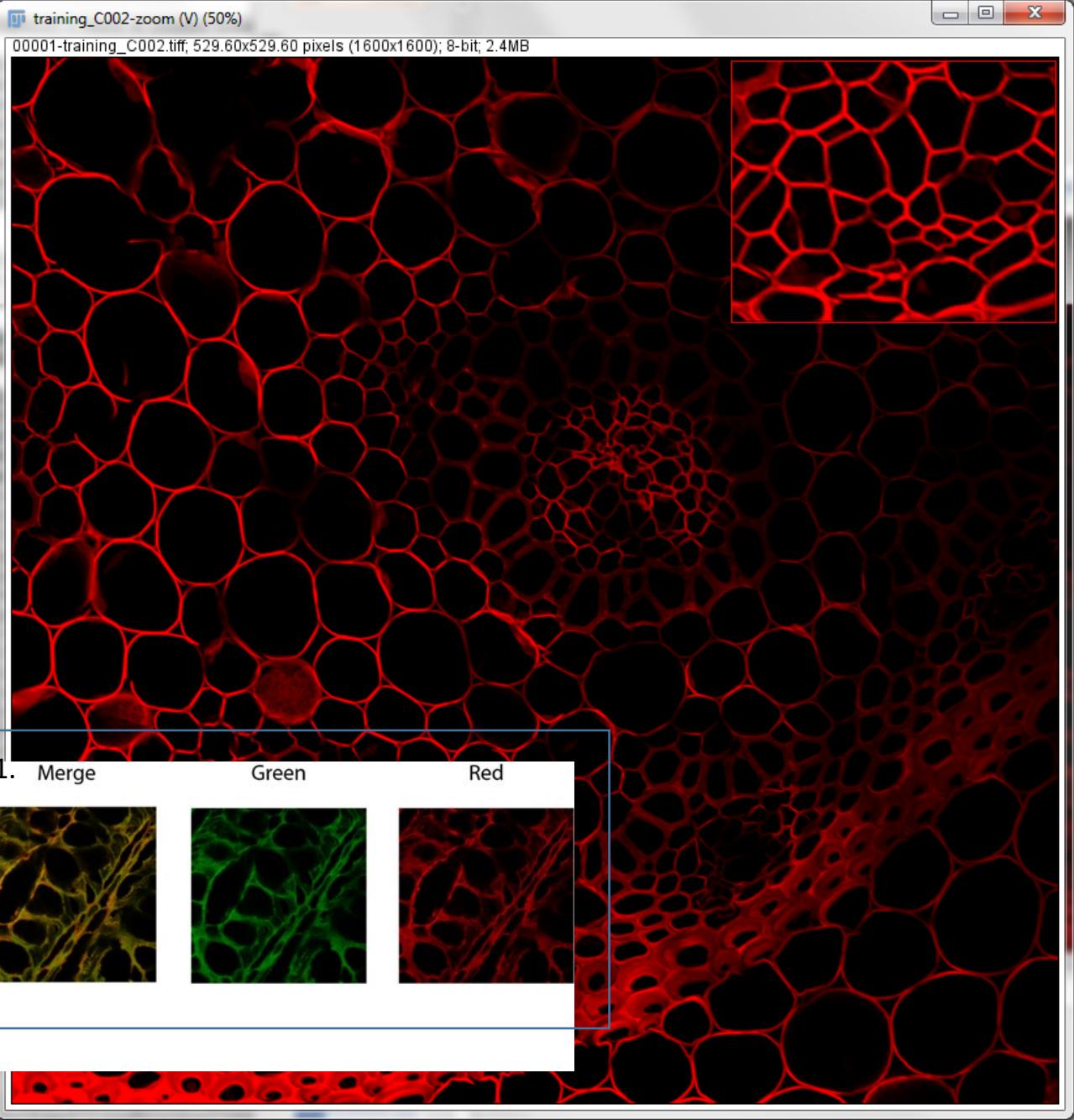
**Imaris**  
3D/4D **Visualization** and  
Analysis Software

Digital Imaging Solutions  
**iTEM**  
Solutions for TEM Applications



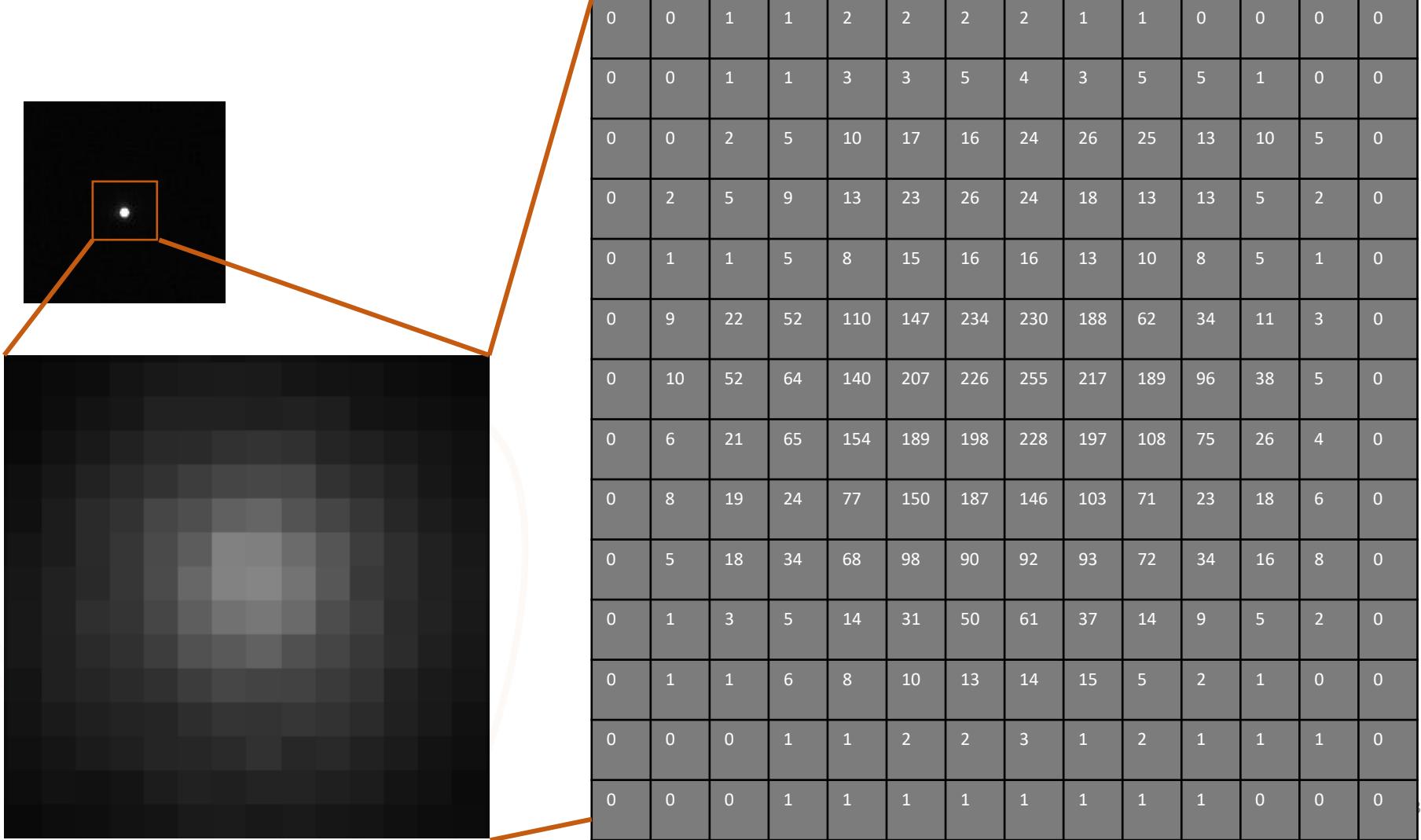
# Outlines

- Basic concepts
- What methods?
- How to process?
- How to analyze?



# What is a digital microscope image?

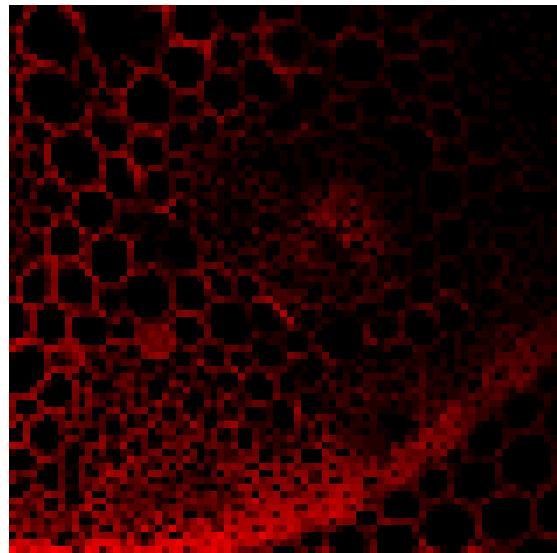
A matrix of pixels



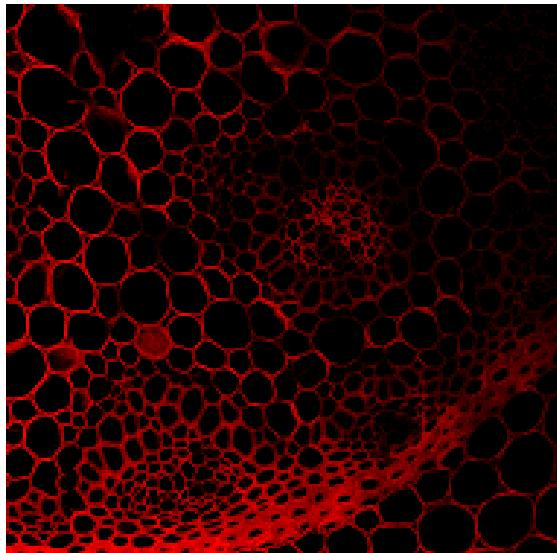
# Image size & image resolution

- Size (dimension)  
in pixels (i.e. 512 X 512)—pixel dimension or  
in inches/ums (i.e. 2.2 um X 2.2 um)—document dimension
- Resolution (pixel density): in DPI/PPI, pixel size (i.e. 50 nm/px )

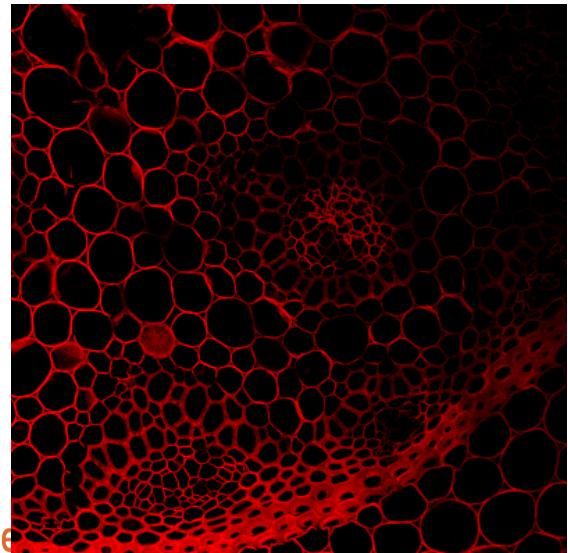
2.67 inches



2.67 inches



2.67 inches



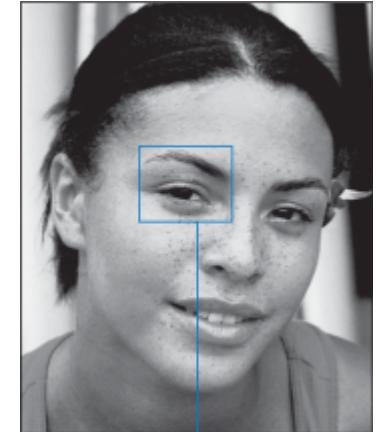
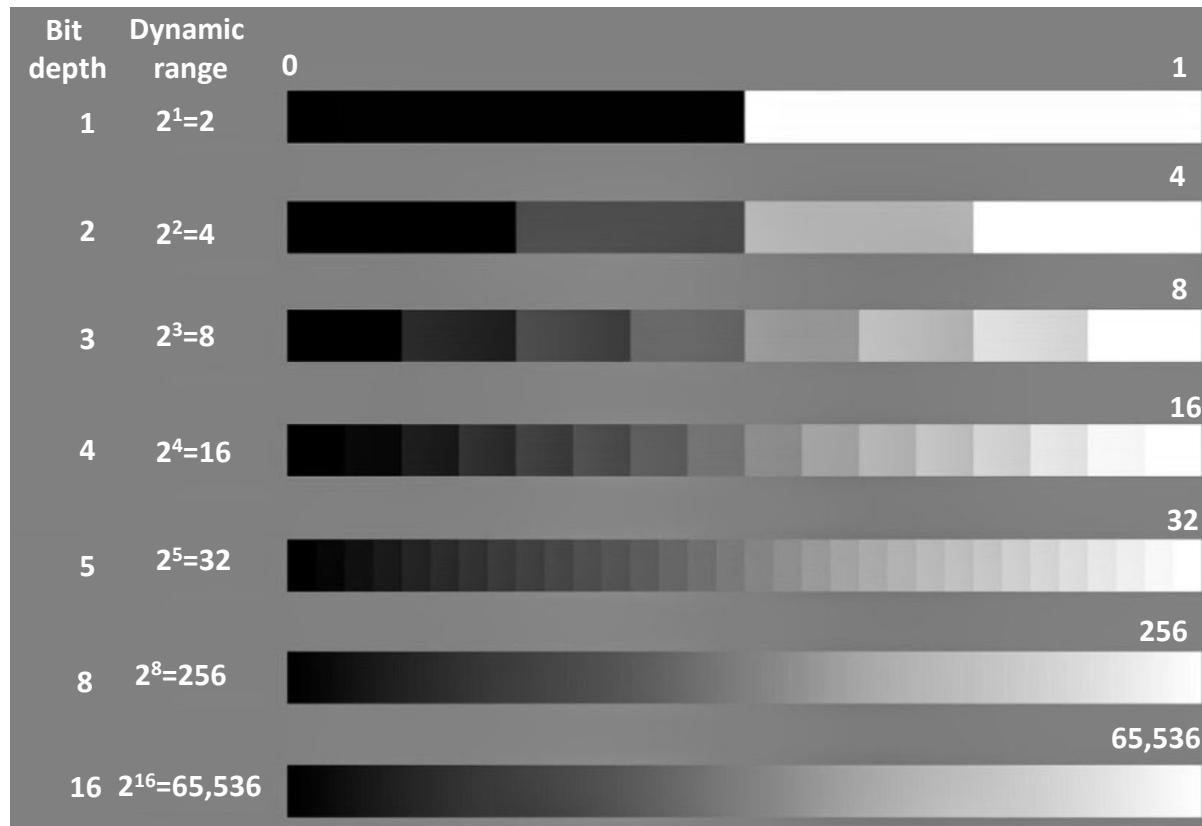
cure

# Bit depth & dynamic range

**Bit**: short for binary digit 0 or 1, smallest unit of intensity data

**Bit depth**: the number of bits. i.e. 8-bit, 00000000-11111111

**Dynamic range**: possible grey shades,  $=2^X$  (X: bit depth)



1 bit



2 bits



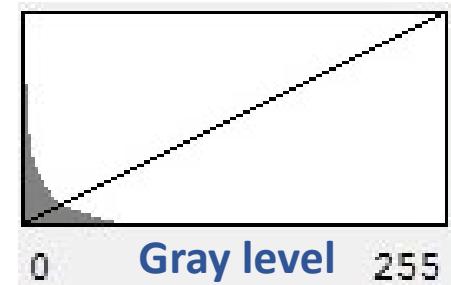
4 bits



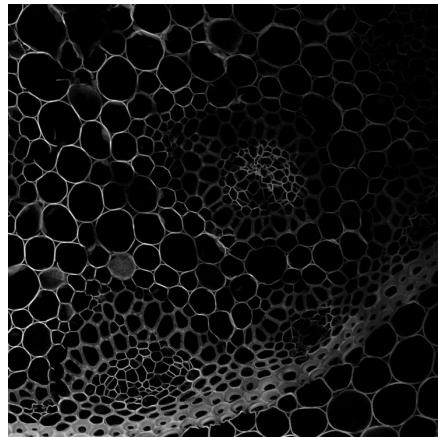
8 bits

# Histogram & manipulation

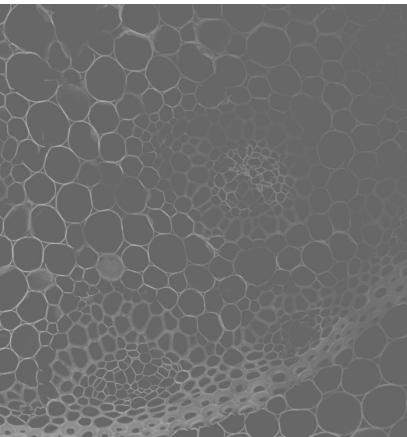
Pixel counts



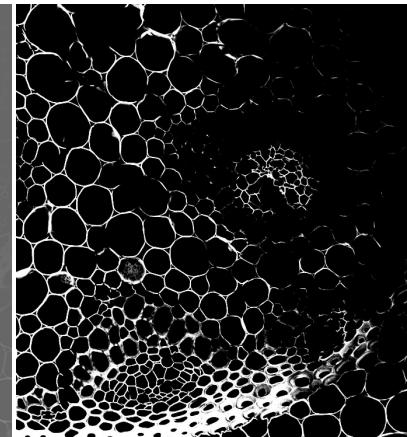
Unprocessed



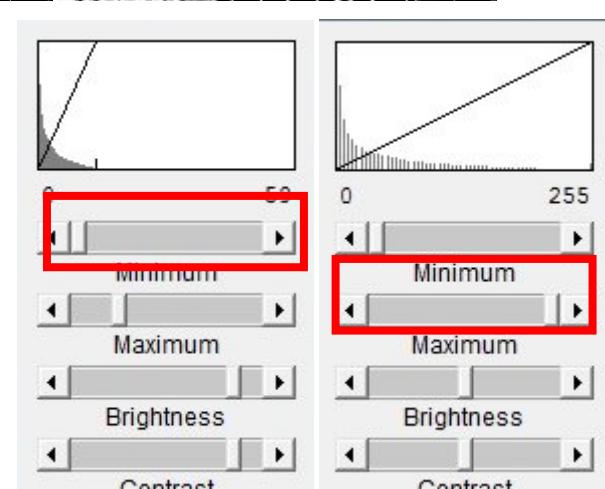
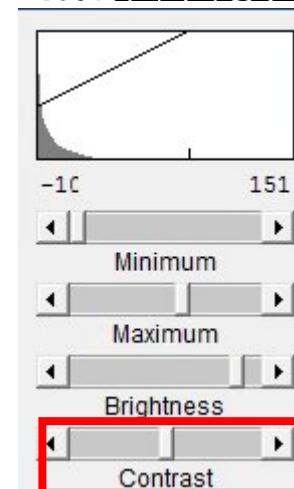
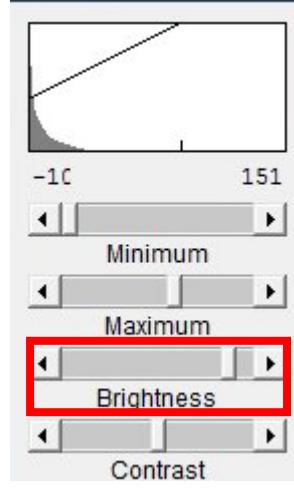
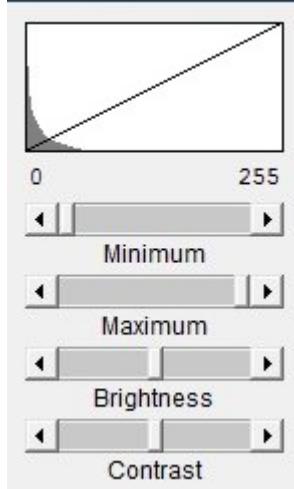
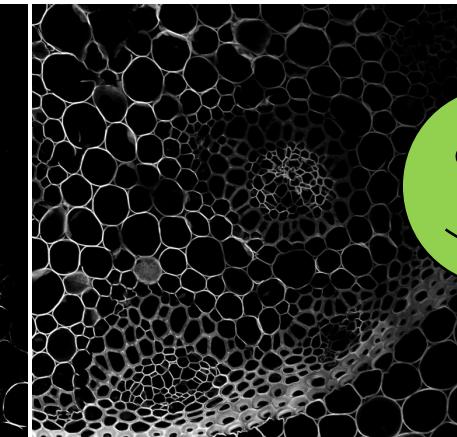
Brightness adjustment



Contrast adjustment

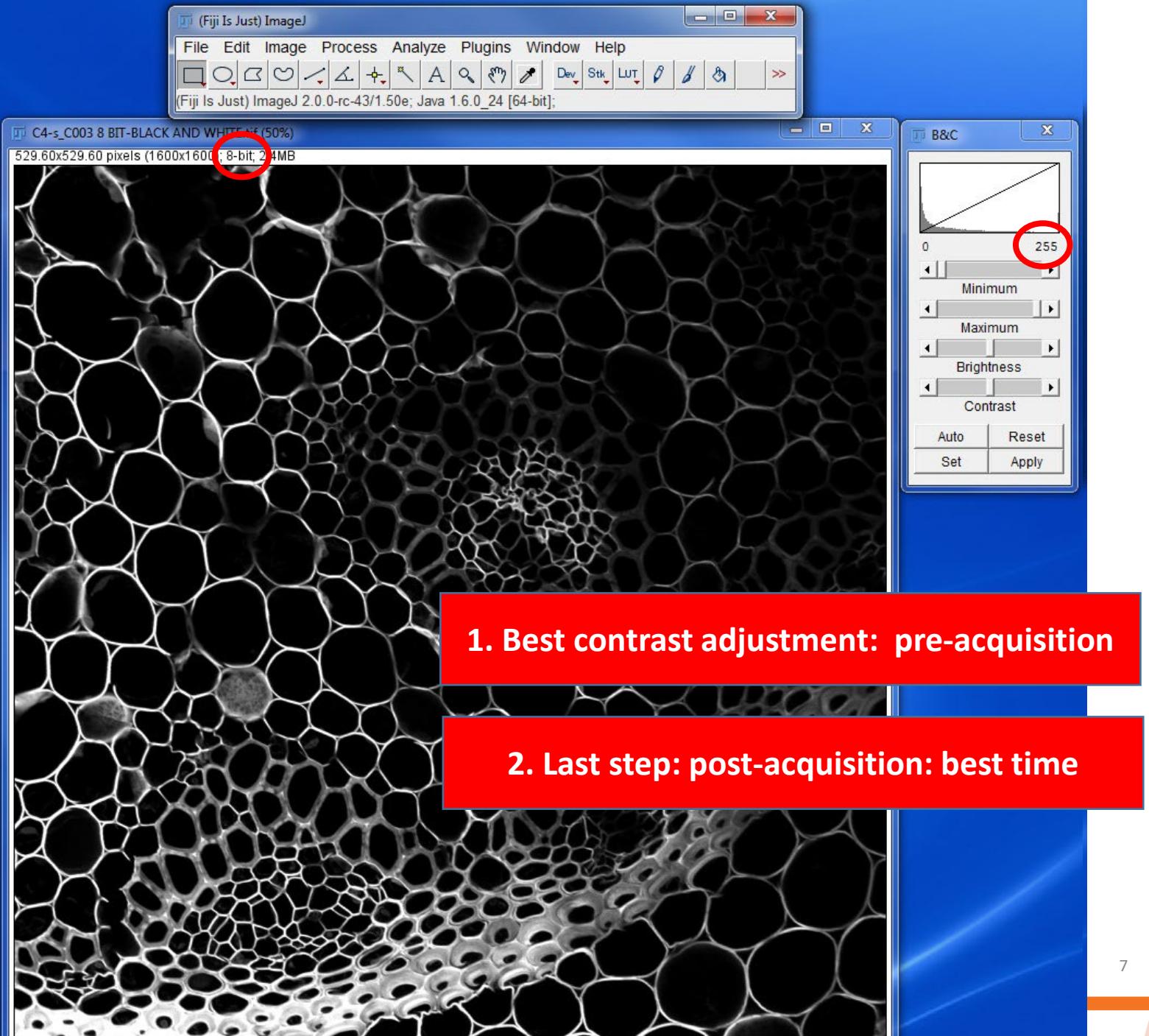
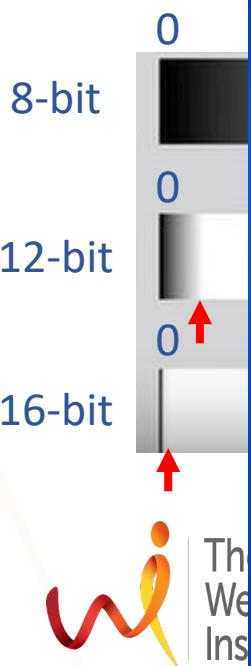


Contrast stretch



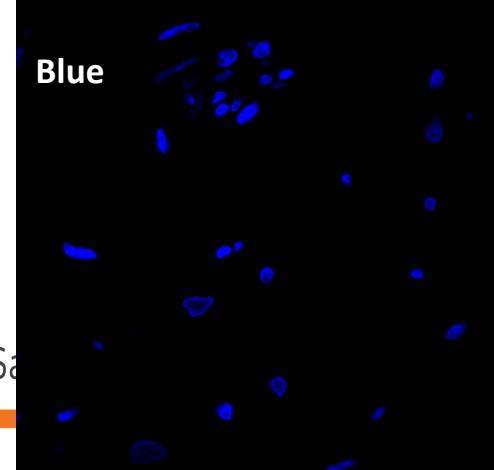
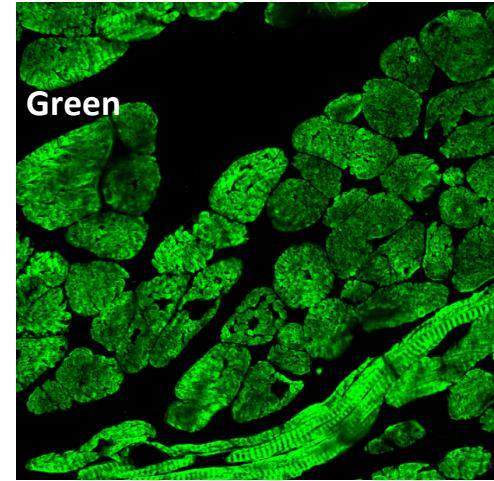
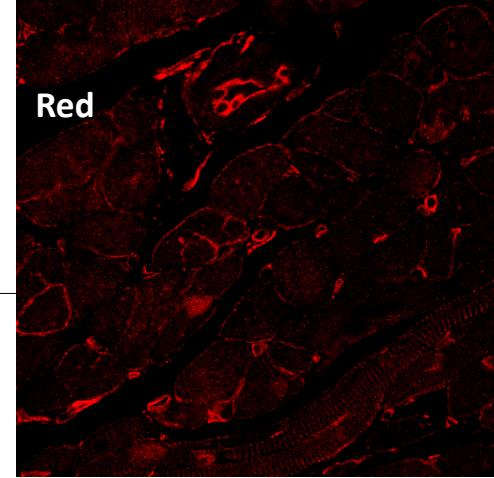
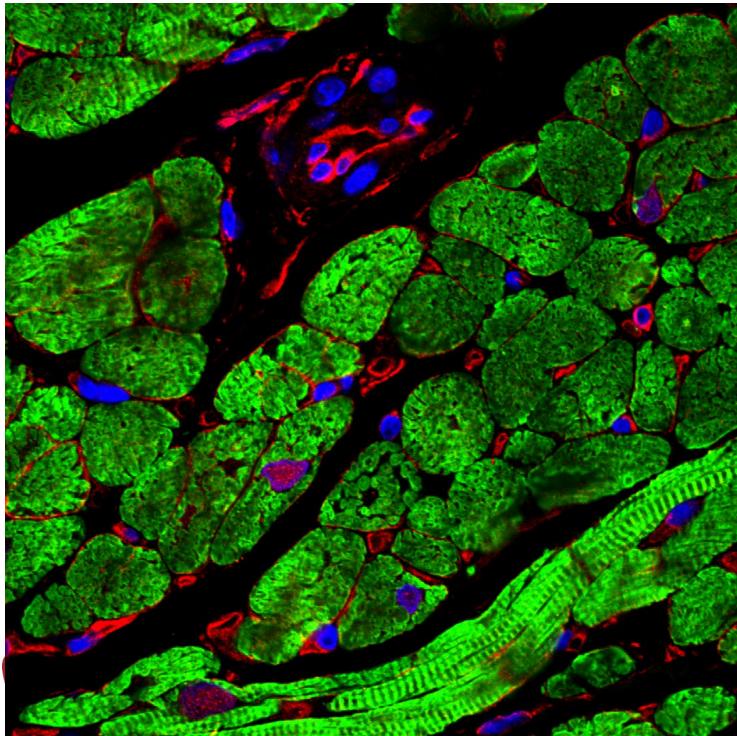
# Conv

- Imaging
- Low sign
- Journals



# Color images

- Simplest color representation is grayscale
- Made up of 3 gray scale channels (RGB)
- Can be 8 or 16 bits per channel (255/65536)



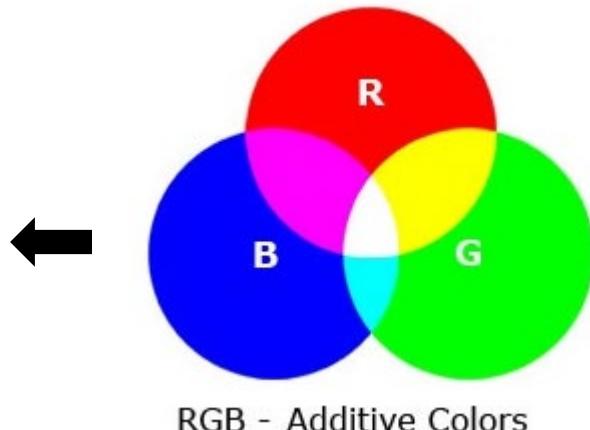
Finding **cures**. Saving **lives**.

# Color models: RGB & CMYK

---

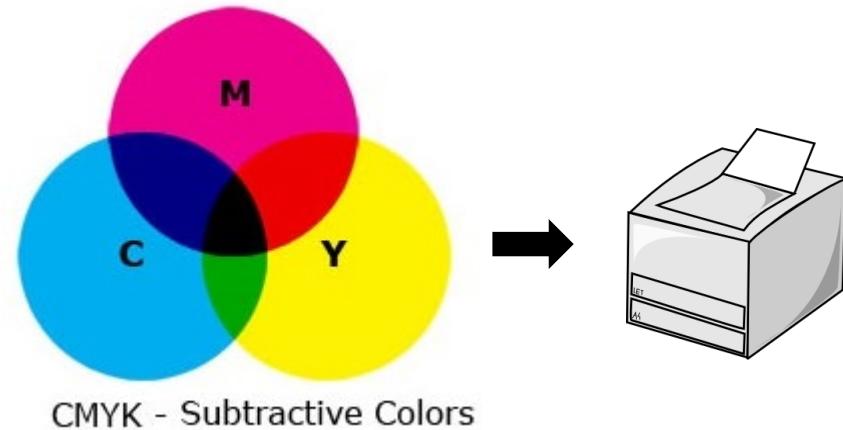
RGB

Red Green Blue



CMYK

Cyan, Magenta, Yellow, Black



Images modified from: <http://inkmonstr.com/2018/02/23/cmyk-vs-rgb/>

# Image formats

---

## The contents of an image file

- Image data: pixel values (numbers, only numbers)
- Metadata: data about data (image type, bit depth, pixel size, microscope settings etc)

## File saving

For analysis: formats best preserving data

Display: general formats



Always keep  
your original  
data!

# Commonly-used general formats

---

Recommended (lossless): **Tiff**

Generally good (lossless): JPEG2000, BMP, PNG

Generally bad (lossy): JPEG, JPG, GIP



Avoid JPEG!

# Bitmap (raster) & vector images

 [scinemag.org/authors/instructions-preparing-revised-manuscript](http://scinemag.org/authors/instructions-preparing-revised-manuscript)

## Format

Figure files at the revision stage must be in one of the following formats (in preferred order):

***Vector illustrations and diagrams (preferred):*** Adobe Portable Document Format (PDF)

Encapsulated PostScript (EPS), or Adobe Illustrator (AI).

***Raster illustrations and diagrams:*** Tagged Image File Format (TIFF)(minimum 300 dpi).

*Vector and raster combinations for photographs or microscopy images:* Adobe Portable Document Format (PDF) or Encapsulated PostScript (EPS)

***Raster photographs or microscopy images:*** Tagged Image File Format (TIFF)

Please keep an archive of all original images used in figures as Science may request delivery of these images for production purposes. Save these at the highest resolution possible, preferably as the original file in its native format.

*At this stage in the process, we cannot accept files in formats other than those specified above; in particular, we cannot accept:*

- Figures embedded in Microsoft Word files.
- Microsoft PowerPoint files.

# What manipulations are “legal”?



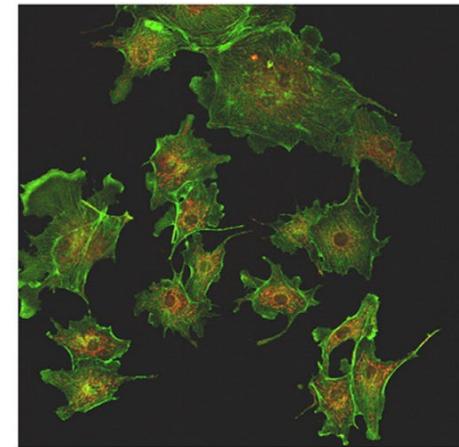
- Linear adjustment of brightness, contrast, color balance in moderation
- Background subtraction
- Cropping
- Reduce image resolution



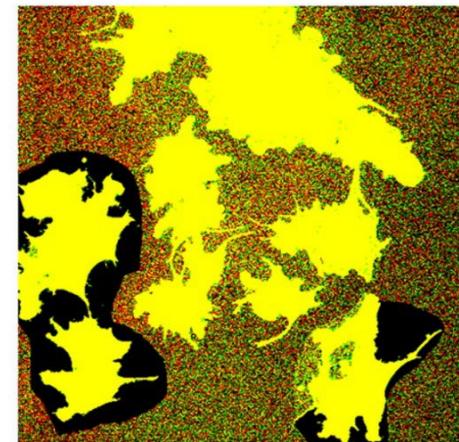
- Increasing image resolution
- Adjusting only a specific part of an image or erasing spots
- No cutting/pasting into a single picture
- Control and experiment are not treated identically



Manipulated image



Manipulation revealed by contrast adjustment



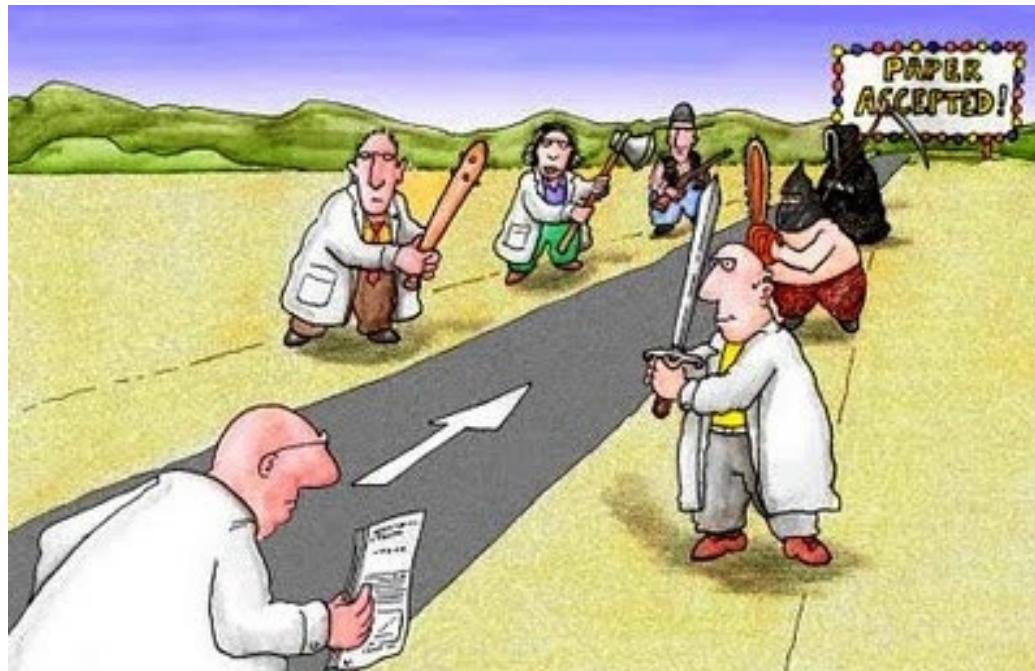
# Suggestions on image manipulations

---

- Keep original data as it was acquired
- Perform adjustments on a copy of the unprocessed image
- Save processed images separately with important processes or adjustments
- Disclose handling software and specific processing
- Do not increase the resolution of an image when exporting
- Ethical guidelines <http://jcb.rupress.org/content/166/1/11>, <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4114110/>

# Figures: increase clarity of data

Meet Journal formatting requirements!



# Figure-making rules

---

- Read the journal instructions **first**:

*Image type: raster/vector, 8 bit, RGB/CMYK*

*Image size (dimensions): 1 (3.5 inch/9cm) or 2 column (7.3 inch/18.5 cm)*

*Image resolution—input>>> output: 300 or 600 or 1200 dpi/ppi?*

*File size (< 5Mb)*

*Format (Tiff, PDF, etc)*

- Be mindful of **acquisition** resolution > 300 dpi
- Don't manipulate images excessively
- Avoid the use of lossy compression (use recommended format)
- Each figure should be submitted as a single file

# Figure-making software tools

We need proper software to

- arrange, lay out, and annotate your images;
- bring in raster images;
- make/draw vector graphics;
- export the final figure.

Commonly used programs:

- **Word**: bad choice
- **Photoshop**: not recommended
- **Powerpoint**: try to avoid
- **Illustrator**: recommended
- Others: Inkscape, InDesign etc



Maintain  
resolution!

# 4-step figure-making workflow: recommended

**2 software tools are involved: Fiji ImageJ & Illustrator**

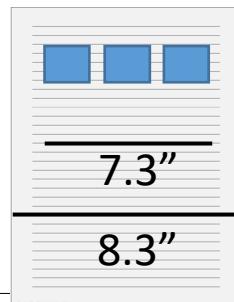


Step 1: Planning: journal requirements, raw data

Step 2: Getting individual images ready: **Fiji ImageJ** (better than Photoshop) size, res, bit depth, format, etc

Step 3. Assembling components: **Illustrator**  
raster & vector images, texts & annotation (vector), etc

Step 4. Export file: **Illustrator**  
resolution (300 DPI), RGB/CMYK, format (PDF/Tiff), etc



# A figure-making example

## Journal requirements

- Single column figure
- Output: 8 bit RGB, 300PPI, Tiff

## Raw images

- Size: 1600X1600 Pixel, 70.356x70.356 um
- Conversion: 1 um = 0.00003937 inch
- Resolution: 1600 pixel/70.356 um (pixel/um)  
 $1600/70.356/0.00003937=577,635 \text{ PPI}$
- Format: oif →→ Tiff (lossless)

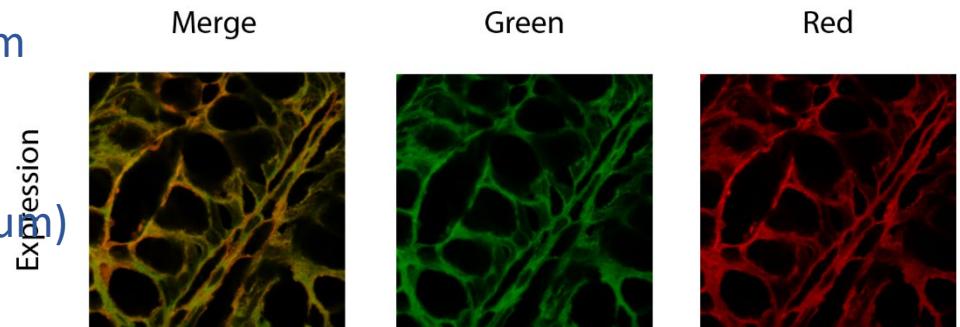
Task: make a figure embedded with 3 fluo images

Plan: A4: 8.3" x 11.7"

single column: 8.3" → → 7.3"

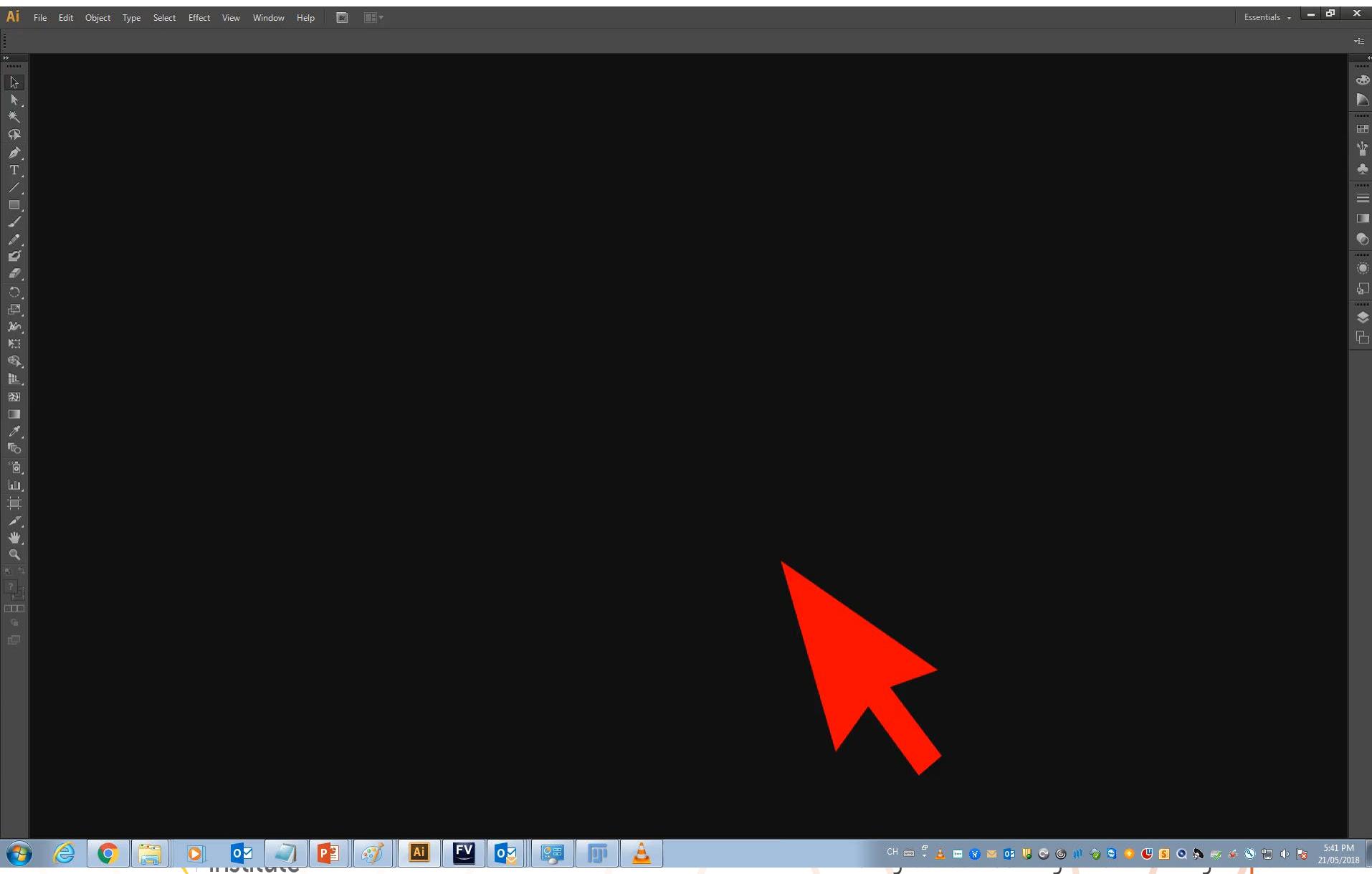
individual channel image:  $7.3"/3=2.4"$   
let's do **2.2"** width!

7.3 inches



Reduce to 300 PPI

2.2 inches



# Why Illustrator not Powerpoint?

sciemag.org/authors/instructions-preparing-revised-manuscript

## Format

Figure files at the revision stage must be in one of the following formats (in preferred order):

*Vector illustrations and diagrams (preferred): Adobe Portable Document Format (PDF)  
Encapsulated PostScript (EPS), or Adobe Illustrator (AI).*

*Raster illustrations and diagrams: Tagged Image File Format (TIFF)(minimum 300 dpi).*

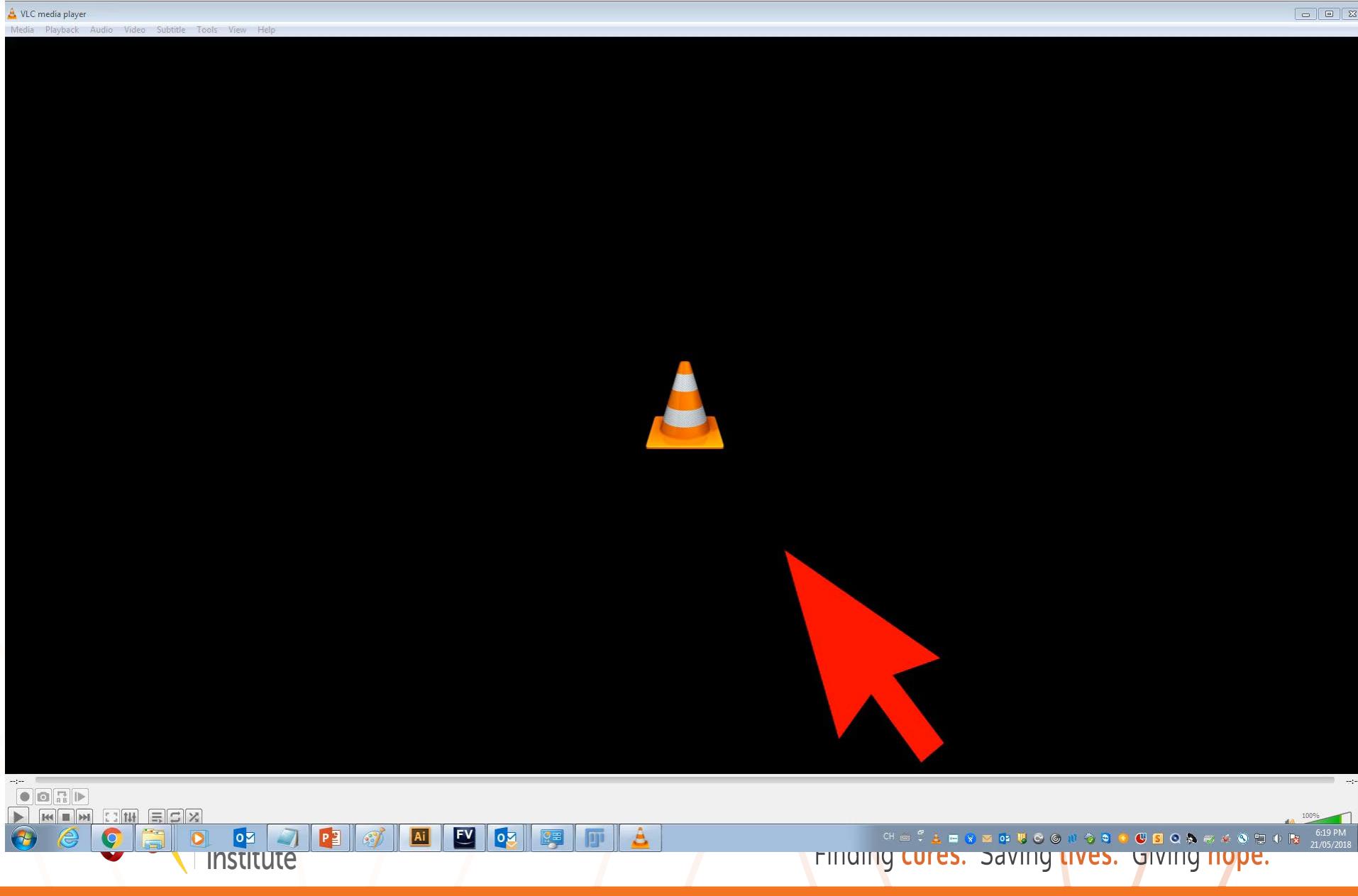
*Vector and raster combinations for photographs or microscopy images: Adobe Portable Document Format (PDF) or Encapsulated PostScript (EPS)*

*Raster photographs or microscopy images: Tagged Image File Format (TIFF)*

Please keep an archive of all original images used in figures as Science may request delivery of these images for production purposes. Save these at the highest resolution possible, preferably as the original file in its native format.

*At this stage in the process, we cannot accept files in formats other than those specified above; in particular we **cannot** accept:*

- Figures embedded in Microsoft Word files.
- Microsoft PowerPoint files.



Cre

1. Open [https://imagej.nih.gov/ij/macros/tools/Zoom\\_in\\_Images\\_and\\_Stacks.txt](https://imagej.nih.gov/ij/macros/tools/Zoom_in_Images_and_Stacks.txt)

2. In FIJI:

- Open the image, select a ROI
- Press "Ctrl+Shift+n"—you'll get an Editor window
- Paste the above copied macros command here
- Click "run"

Paste commands here

```

114 dialog.addcheckbox("outline destination", SHOWDESTINATION);
115 Dialog.addNumber("Line width:", surZoom, 0, 1, "");
116 if (slices > 1) {
117     Dialog.addMessage("");
118     fromSlice=1; toSlice=slices;
119     Dialog.addNumber("First slice:", fromSlice, 0, 4, "");
120     Dialog.addNumber("Last slice:", toSlice, 0, 4, "");
121 }
122 Dialog.show();
123 zoomValue = Dialog.getNumber();
124 showInitialSelection = Dialog.getCheckbox();
125 surOri= Dialog.getNumber();
126 showInitialSel = Dialog.getCheckbox();
127 surZoom= Dialog.getNumber();
128 if(slices > 0) {
129     fromSlice= Dialog.getNumber(); FSlice=parseFloat(fromSlice);
130     toSlice= Dialog.getNumber(); Tslice=parseFloat(toSlice);
131 }
132 if (zoomValue < 1) zoomValue =1;
133 if ((widthSel * zoomValue) >= width || (heightSel * zoomValue) >= height) {ok=0;} else {ok=1;
134 }
135 }
136

```

Choose settings

Zoom factor: 2.0

Outline source

Line width: 1

Outline destination

Line width: 2

OK Cancel

Run Kill Show Errors Clear

Overlay.drawRect(xz0...

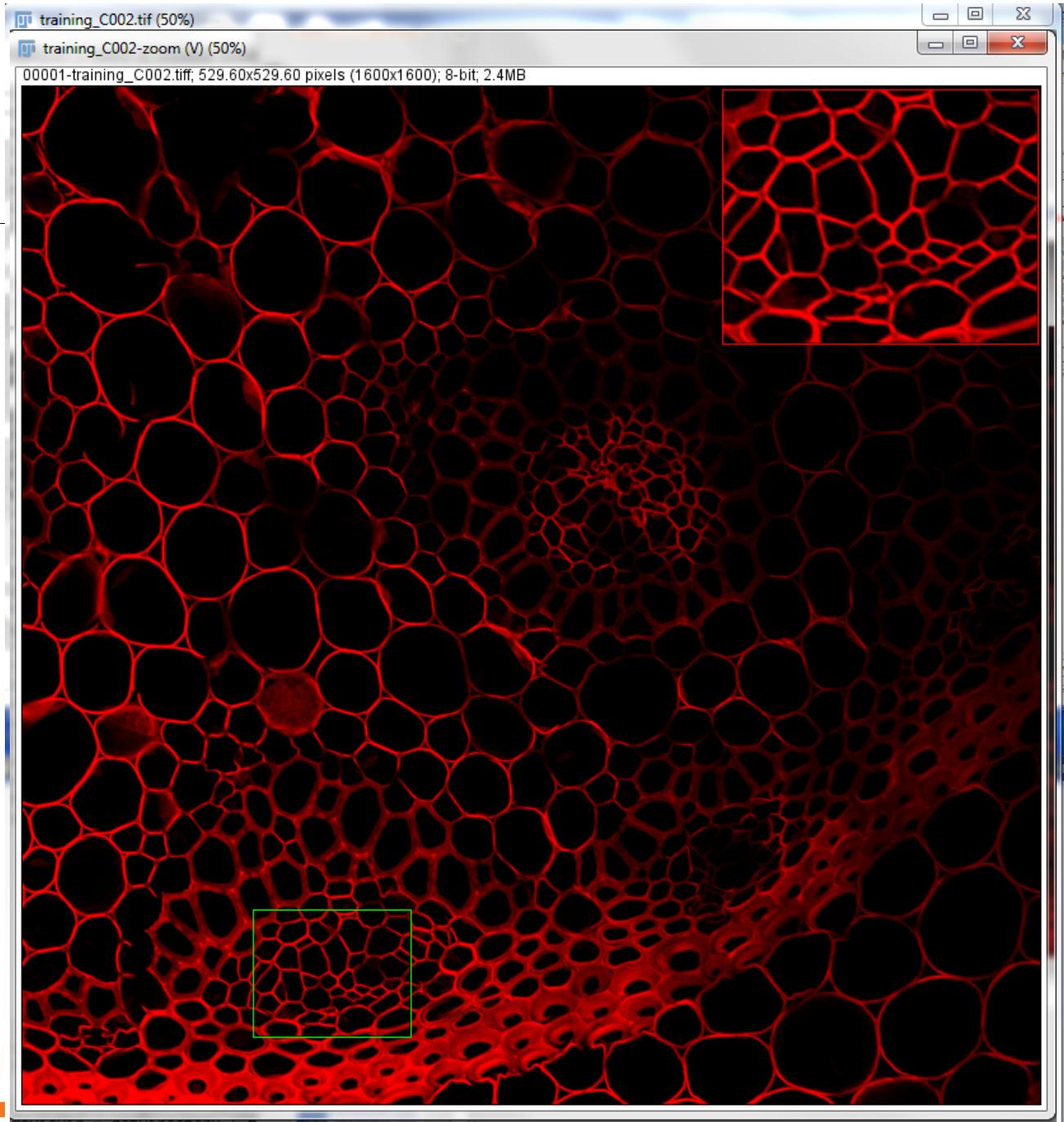
Speed up browsing by disabling add-ons.

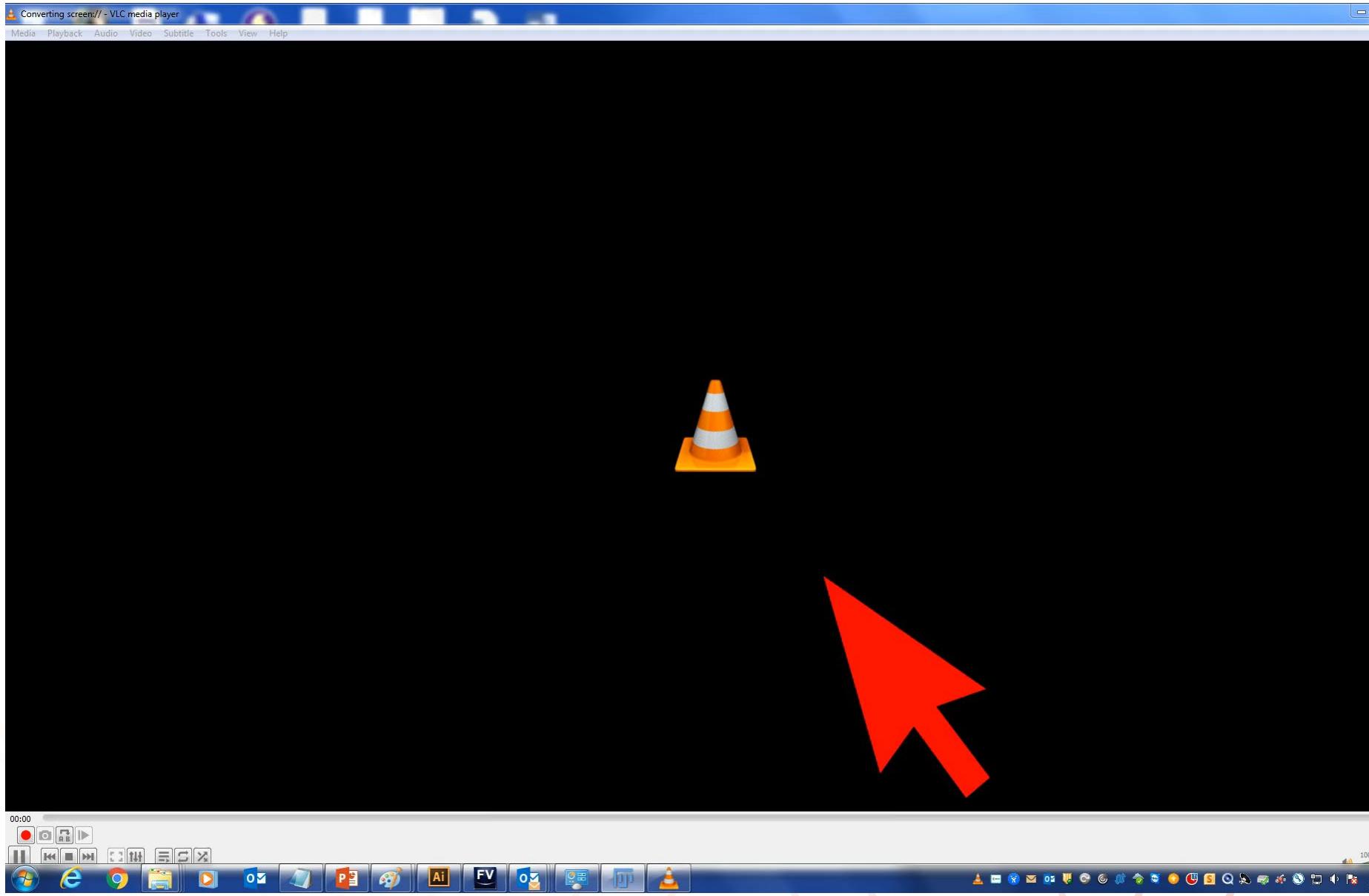
24

# Creating an

4. Now you decide where to place your inset (red square)

5. Finally click on the red square place the inset





# Summary & take-home messages

---



# Available image processing & analysis software

---

1. Fiji ImageJ (general, open source)
2. Aivia: general (analysis, DPL)
3. Desktop 200: general (analysis, DPL)
4. Others: ASW (Olympus VS 120), LASX (Leica), Zen (Zeiss), NDP view2 (NZ), Imagescope (Leica)

# Visit Our website to find this presentation...

<https://sydneyuni.atlassian.net/wiki/spaces/WIF/pages/765397549/Tips+Tricks>

<https://wimr.sharepoint.com/sites/ScientificPlatform2/SitePages/Imaging%20Facility.aspx>





# Thank you!