

Image Processing and Analysis

Time	Presentation	Speaker
1.00-1.10 pm	Welcome	Hong Yu/WIMR
1.10-1.45 pm	Things you need to know for publishing images	Hong Yu/WIMR
1.45-2.20 pm	A brief introduction to image analysis using ImageJ	Josh Studdert/CMRI
2.20-2.55 pm	An introduction to deconvolution with Huygens	Laurence Cantrill/KRI
2.55-3.15 pm	Afternoon tea	Hong Yu/WIMR
3.15-3.50 pm	Multi-dimensional image analysis using Imaris	Scott Page/CMRI
3.50-4.25 pm	Image analysis in Electron Microscopy	Emma Kettle WIMR/CMRI
4.25-4.30 pm	Wrap up + upcoming events	Hong Yu/WIMR

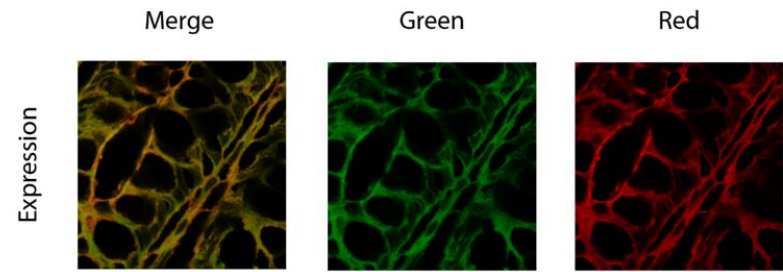
Finding **cures**. Saving **lives**. Giving **hope**.



Things you need to know about figure-making for final publishing

Hong Yu, Cell Imaging Facility 24 May 2018

Introduction



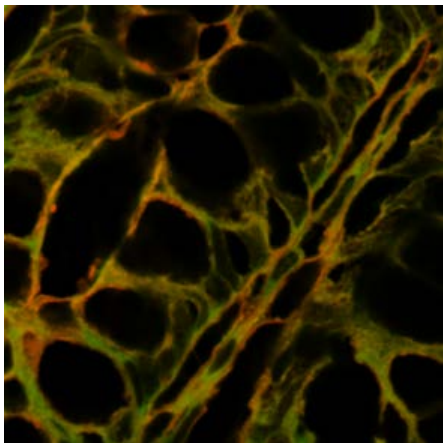
Raw
images

Processing

Analysis

Figure

Publication



Imaris
3D/4D **Visualization** and
Analysis Software

Digital Imaging Solutions

iTEM

Solutions for TEM Applications



Images modified from <https://fiji.sc/>; <https://svi.nl/HomePage>; <http://www.sciencemag.org/topic/optics>; <https://science.institut-curie.org/platforms/cell-and-tissue-imaging/photronics/pict-lm-software/imaris/>; <https://www.behance.net/gallery/8956263/Nature-science-journal-cover>
http://photobucket.com/gallery/http://s831.photobucket.com/user/hibari_sensei/media/cellcover_zps342b968d.jpg.html

Outlines

- **Important concepts and terms**
- **What manipulations are “legal”?**
- **How to make figures for journals?**

What is a digital image? A matrix of pixels

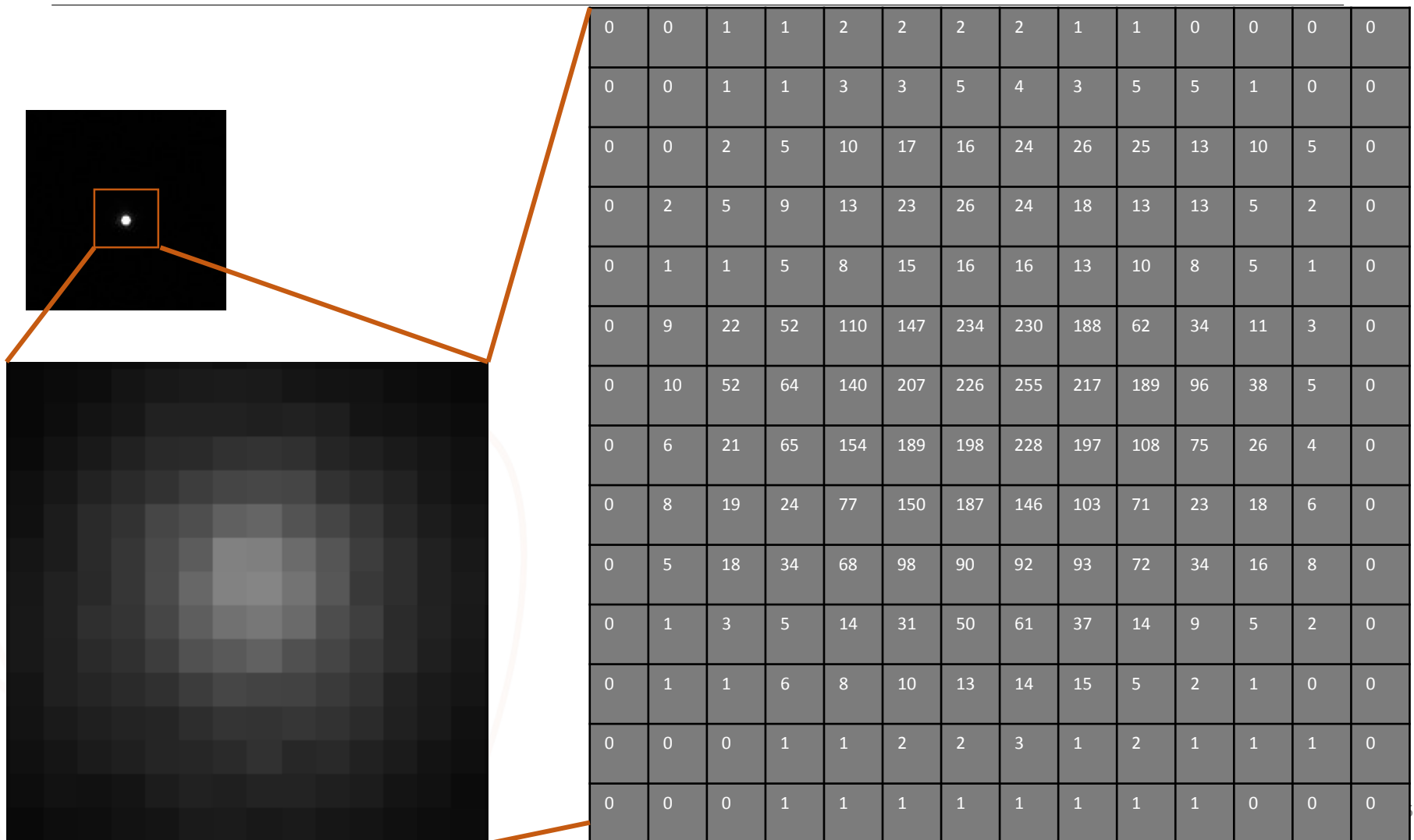
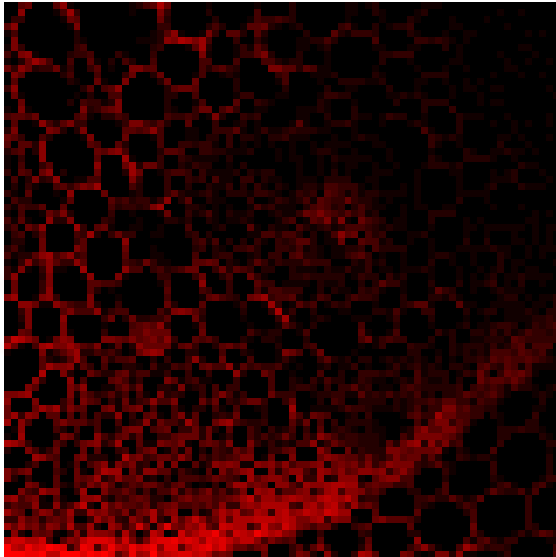


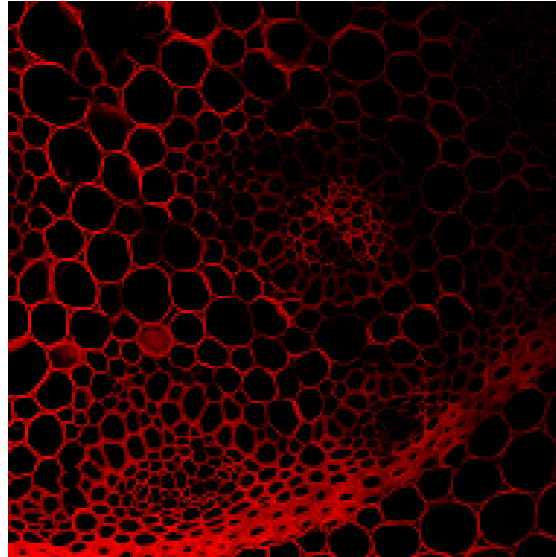
Image size and resolution

- Image size (pixel dimensions): 1024 X 1024
- Image resolution (pixel density):
dpi (dots per inch) or ppi (pixels per inch)

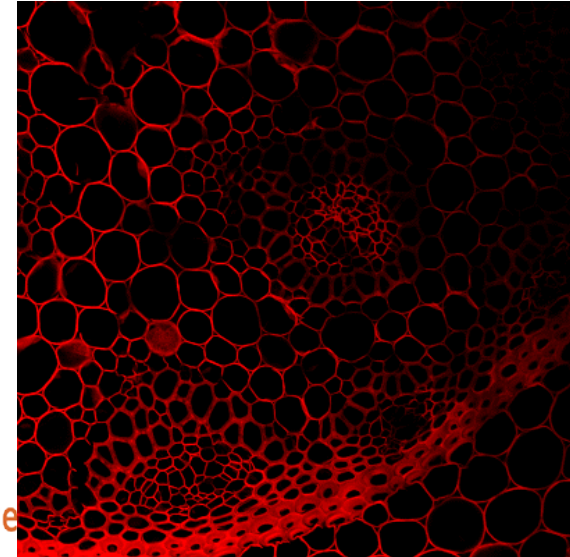
PPI: 30 80X80 6K



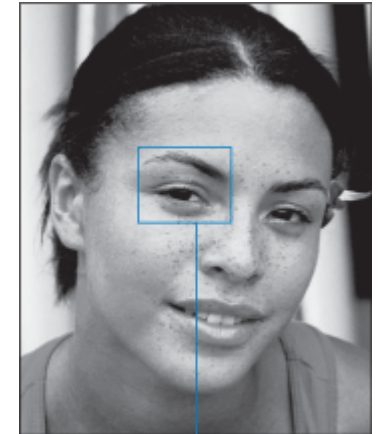
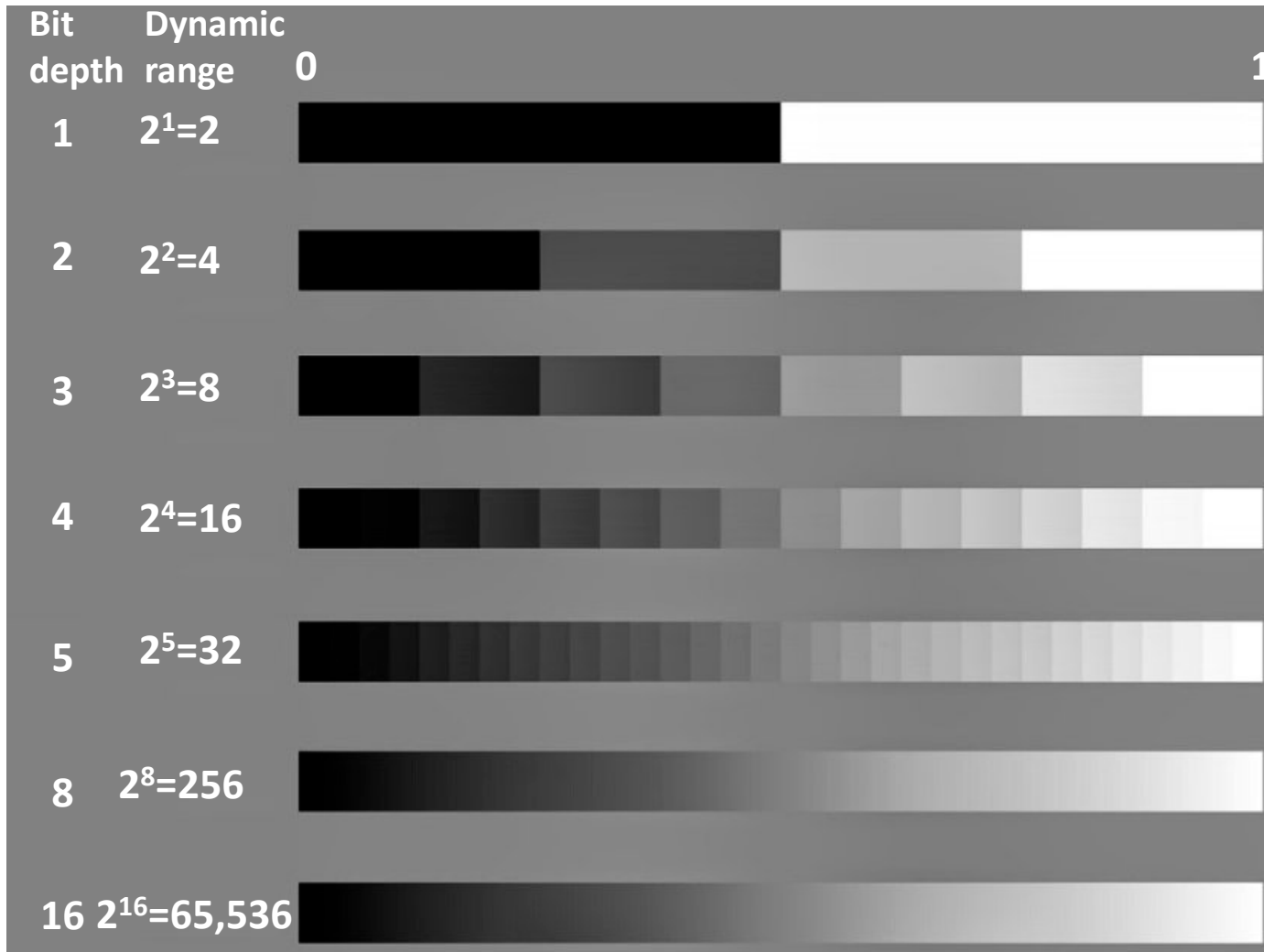
PPI: 72 192X192 36K



PPI: 150 400X400 156k



Bit depth and dynamic range



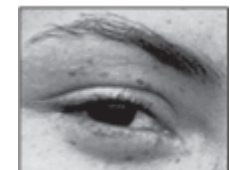
1 bit



2 bits

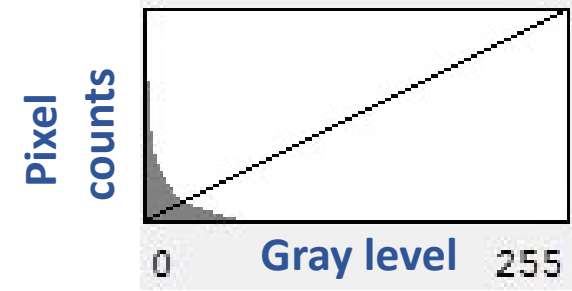


4 bits



8 bits

Manipulating image histogram

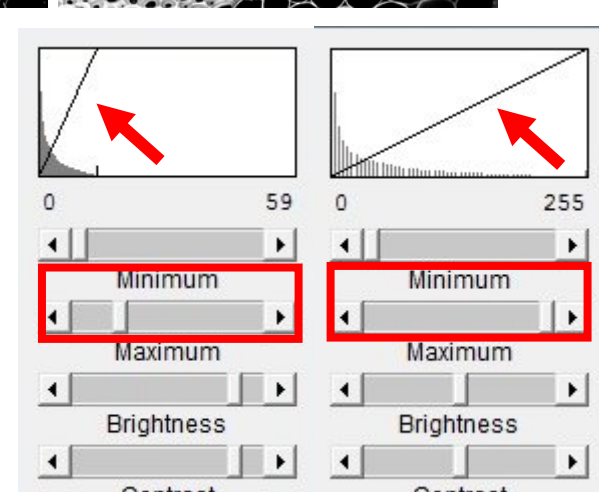
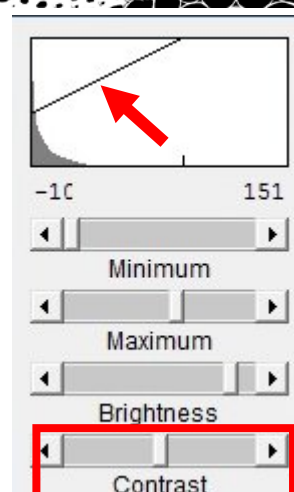
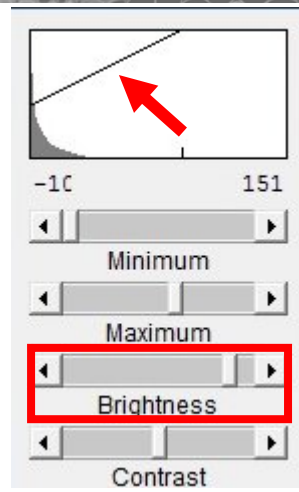
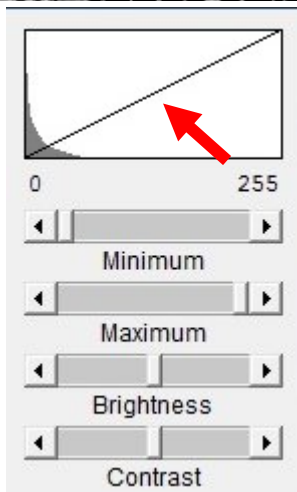
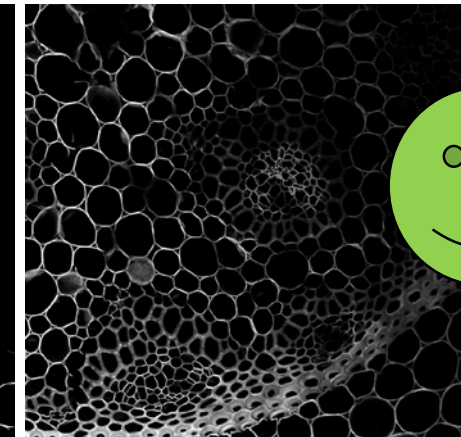
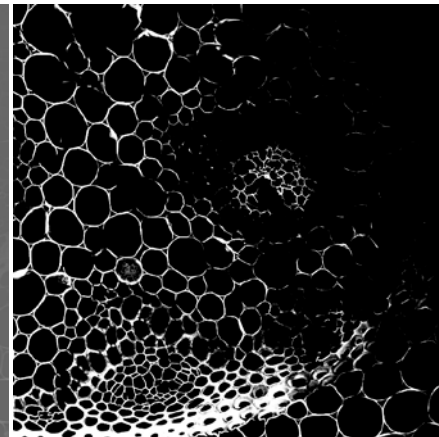
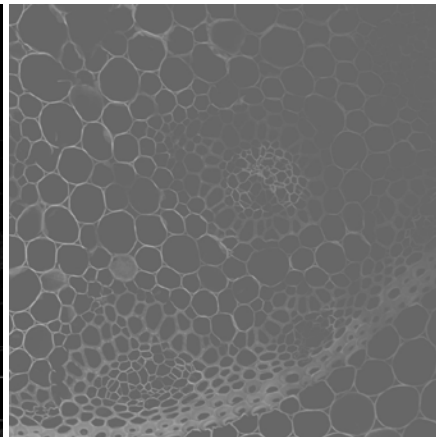
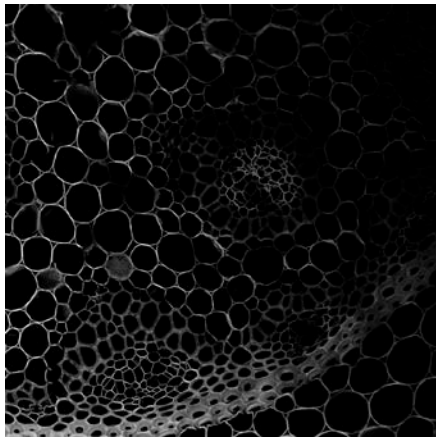


Unprocessed

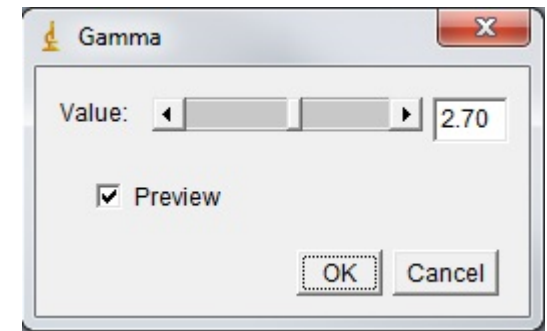
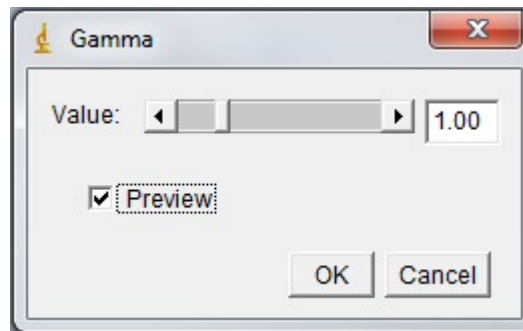
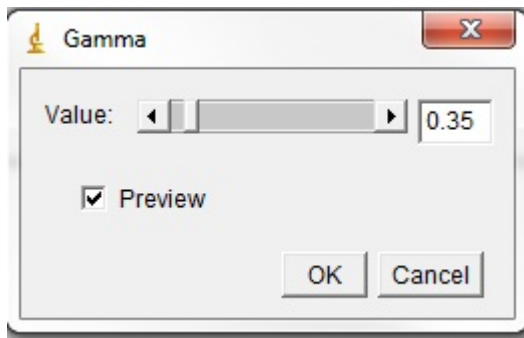
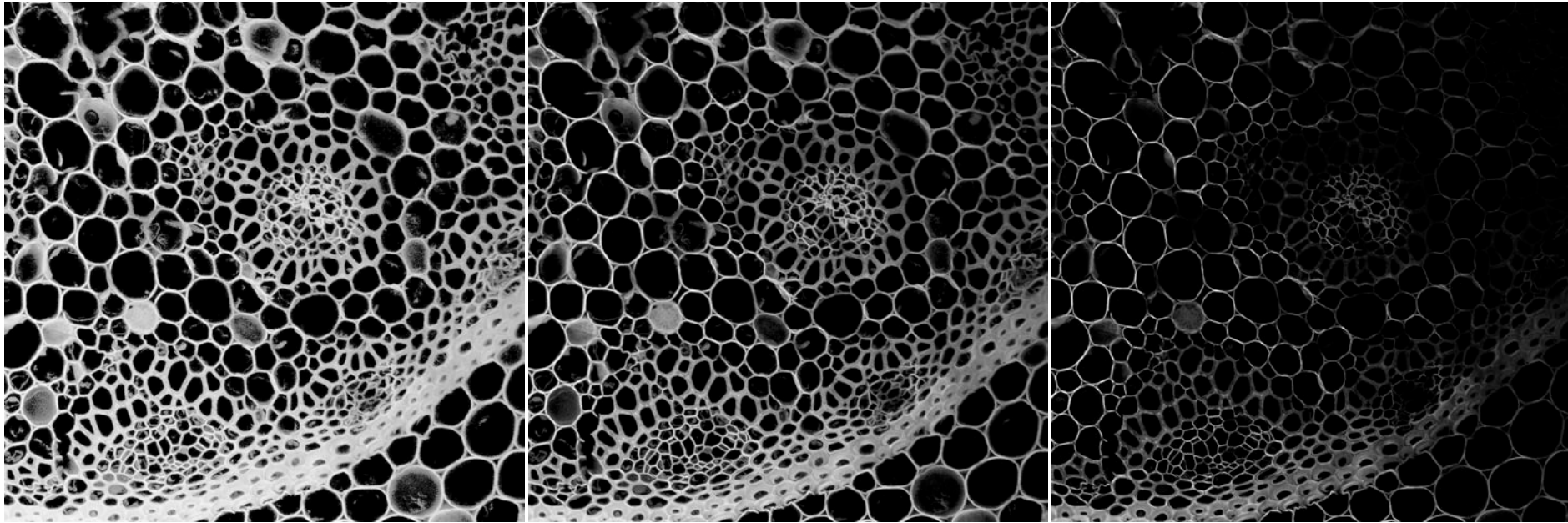
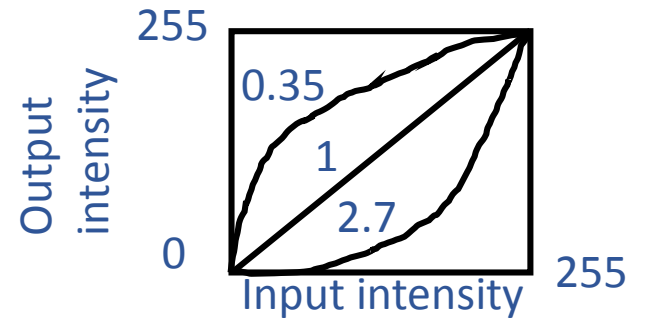
Brightness adjustment

Contrast adjustment

Contrast stretch

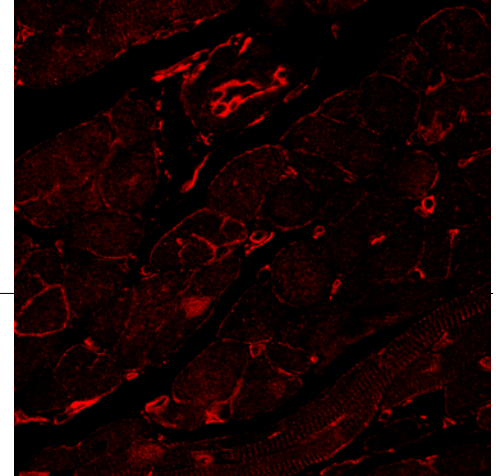


Gamma adjustment



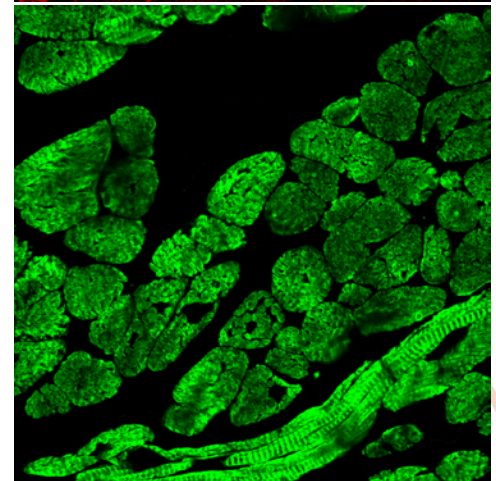
Color images

Red

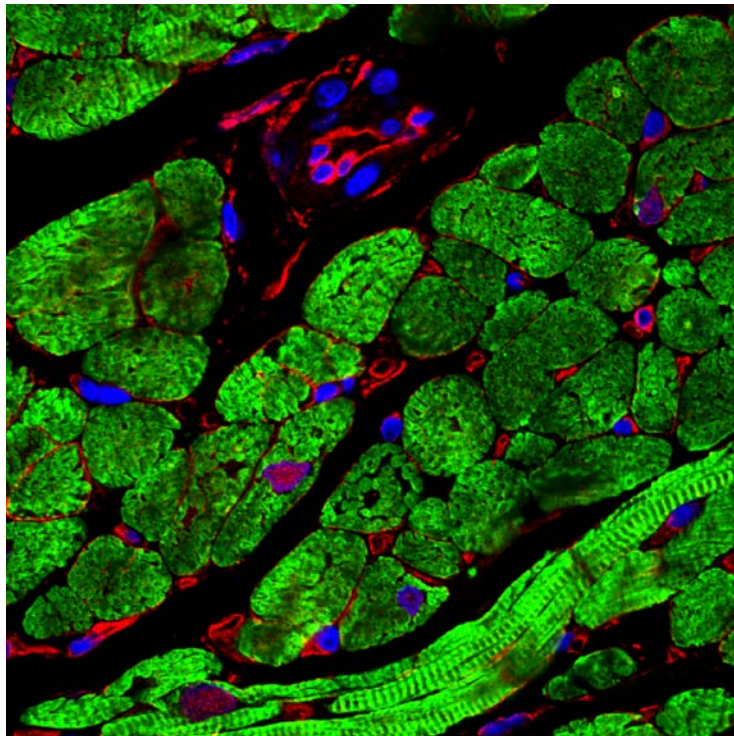
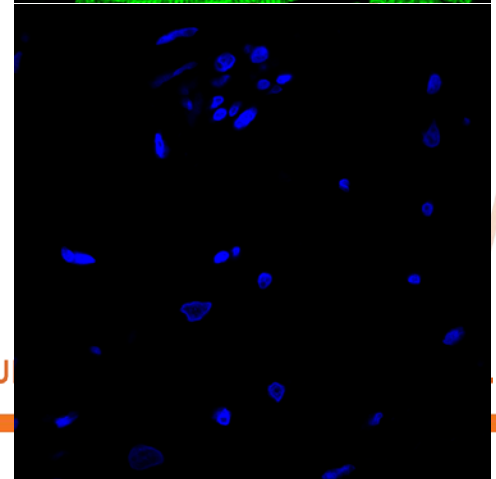


- Made up of 3 gray scale images
- Can be 8 or 16 bits per channel

Green

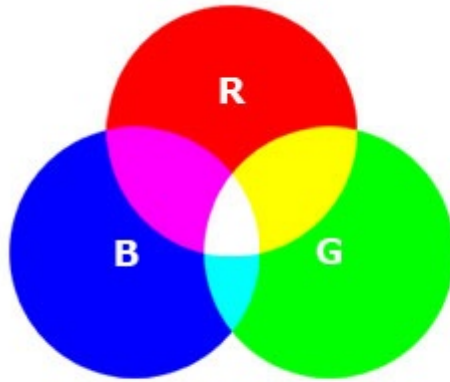


Blue

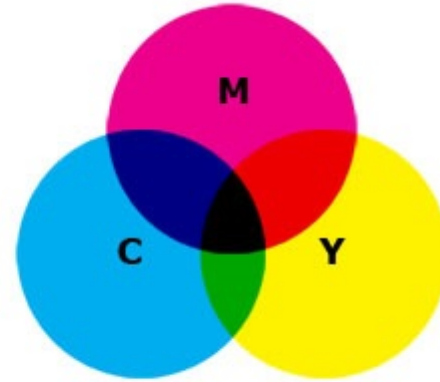


Finding cu

Color models: RGB & CMYK



RGB - Additive Colors



CMYK - Subtractive Colors



Red
Green
Blue

Cyan
Magenta
Yellow
Black

Image formats

The contents of an image file

- Image data: pixel values (numbers, only numbers)
- Metadata: data about data (image type, bit depth, pixel size, microscope settings etc)

File saving

For analysis: formats preserving the metadata

Display: general formats



**Always keep
your original
data!**

Image formats: compression & general formats

Compression

- Lossy: JPEG, GIF, etc
- Lossless: Tiff, JPEG2000



Commonly used formats

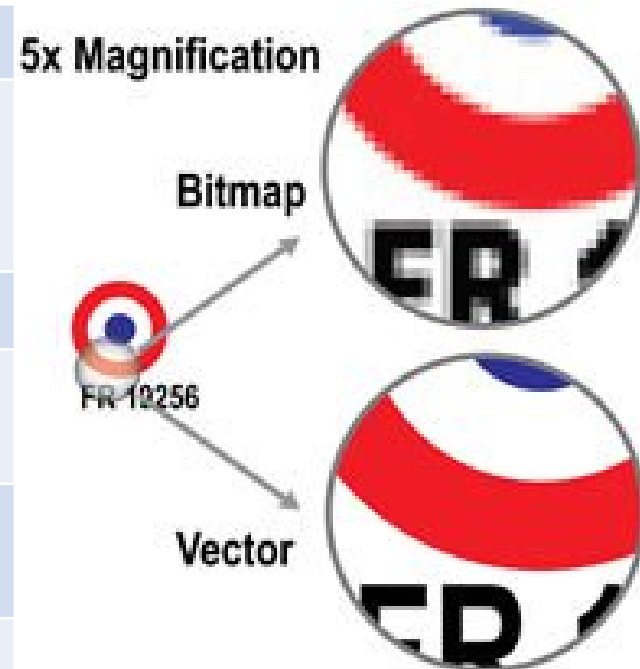
Recommended: **Tiff**

Generally good: **Tiff**, OME-Tiff, JPEG2000, BMP, PNG

Generally bad: JPEG, JPG, GIF

Bitmap & Vector images

	Bitmap/Raster	Vector
Made up of	pixels	paths
Produced by	digital image capture devices (true images)	drawing softwares: illustrator, ppt (math formula)
Example formats	jpg, tiff, bmp	Ai, cdr, dwg
Good used for	color images, mic images	Logos, texts
When enlarged	may appear pixelated	no resolution impact
Resolution	PPI/DPI. Res-dependant	res-independent. Res of output device
Image size	large	small



What manipulations are “legal”?

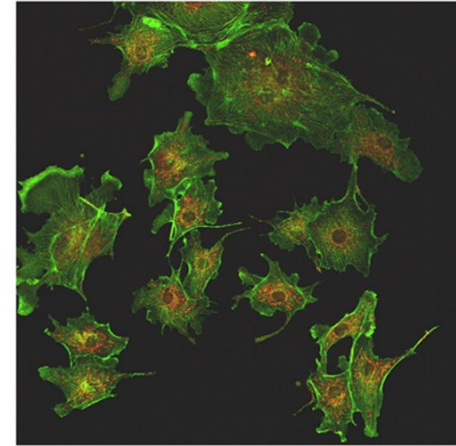


- Linear adjustment of brightness, contrast, color balance in moderation
- Background subtraction
- Cropping
- Reduce image resolution

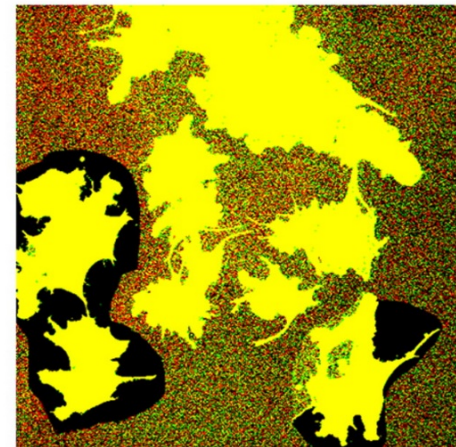


- Increasing image resolution
- Adjusting only a specific part of an image or erasing spots
- No cutting/pasting into a single picture
- Control and experiment are not treated identically

Manipulated image



Manipulation revealed by contrast adjustment



Suggestions on image manipulations

- Safe-keep original data as it was acquired
- Perform adjustments on a copy of the unprocessed image
- Save processed images separately with important process or adjustment
- Disclose handling softwares and specific processing
- Do not increase the resolution of an image when exporting
- Ethical guidelines <http://jcb.rupress.org/content/166/1/11>
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4114110/>

Why building figures?

- Increase clarity of data
- Each figure should be submitted as a single file
- Meet journal formatting requirements

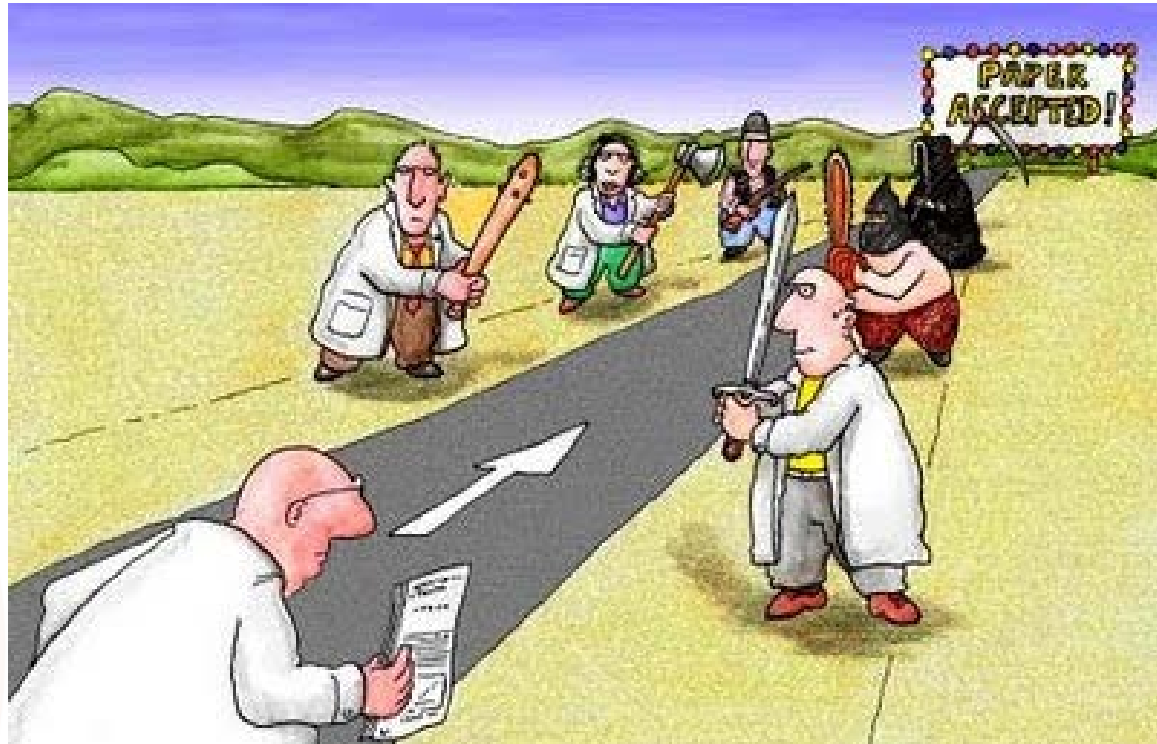


Image courtesy: <http://tripleed.com/lindberg-rantatalo-hallgren-has-article-accepted-on-making-sense-of-paradoxes/>

Figure making: rules

- Read the journal instructions first:

Image type: raster/vector, 8 bit, RGB

Image size (dimensions): 1 (3.5 inch/9cm) or 2 column (7.3 inch/18.5 cm)?

Image resolution—start and final steps: 300 or 600 or 1200 dpi/ppi?

File size (< 5Mb)

Format (Tiff, PDF, etc)

- Be mindful of acquisition resolution > 300 dpi
- Only manipulate images using image manipulation software
- Don't manipulate images excessively
- Avoid the use of lossy compression

Figure making: software tools

We need a proper software to

- arrange, lay out, and annotate your images,
- bring in raster images;
- make/draw vector graphics;
- export the final figure



**Maintain
resolution!**

Commonly used programs:

- Word: bad choice
- Photoshop: not recommended
- Powerpoint: try to avoid
- Illustrator: recommended
- Others: Inkscape, InDesign etc

<http://www.sciencemag.org/site/feature/contribinfo/prep/figguide.pdf>

Figure making workflow---3 steps

After you process and analyse your original images...

1. ImageJ (better than Photoshop): size, res, bit depth
2. Illustrator: assemble vector and raster components
3. Illustrator: export the final figure file:
RGB/CMYK, vector/raster, format, resolution etc

Figure making: an example

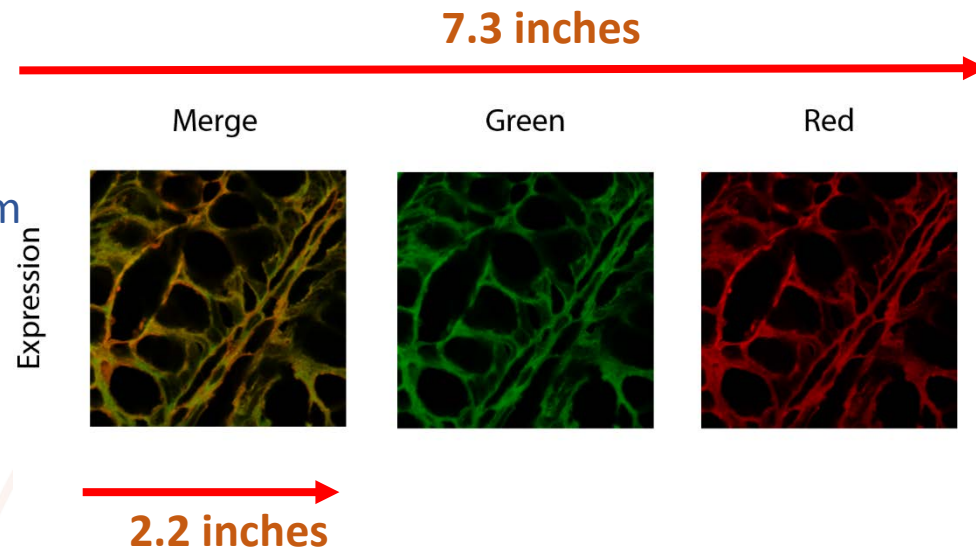
Journal requirements

- Double column figure: 7.3 inches/18.5 cm wide
- Output: 8 bit RGB, 300PPI, Tiff

3 fluo images & texts
each image width: $7.3/3=2.4$
let's do 2.2 inch width

To be assembled images

- Size: 1600X1600 Pixel, 70.356x70.356 μm
- Conversion: 1 μm = 0.02 inch
- Acquisition resolution
 $1600/70.356/0.02=1137$ PPI





Why Illustrator not Powerpoint?

	Powerpoint	Illustrator
Image resolution (new file)	No control	Controllable
Image resolution (export)	Default: 72 DPI < 300 DPI	Controllable
White margin	No control	Auto adjust
Meet journals requirements	May not*	Yes
Good for	Presentation	Printing

* <http://www.sciencemag.org/site/feature/contribinfo/prep/figguide.pdf>



Cre

1. Open up [https://ima](https://imagej.nih.gov/ij/macros/tools/Zoom_in_Images_and_Stacks.txt)

2. In FIJI:

- Open th
- Press “
- Paste t
- Click “r

The screenshot shows a web browser window with the URL https://imagej.nih.gov/ij/macros/tools/Zoom_in_Images_and_Stacks.txt highlighted in a red box. Below the browser is the FIJI software interface. The main window displays a macro script for zooming in on images and stacks. A red arrow points to the line `Dialog.addNumber("Line width:", surZoom, 0, 1, "");` with the text "Paste commands here" next to it. A red circle highlights the "Run" button at the bottom of the macro editor. In the bottom-left corner, a "Choose Settings" dialog box is open, showing options for "Zoom factor" (set to 2.0), "Outline source" (checked), "Line width" (set to 1), "Outline destination" (checked), and "Line width" (set to 2). The "Run" button in the dialog is also highlighted with a red circle.

```
Dialog.addCheckbox("Outline destination", showDestination);
115 Dialog.addNumber("Line width:", surZoom, 0, 1, "");
116 if (slices > 1) {
117     Dialog.addMessage("");
118     fromSlice=1; toSlice=slices;
119     Dialog.addNumber("First slice:", fromSlice, 0, 4, "");
120     Dialog.addNumber("Last slice:", toSlice, 0, 4, "");
121 }
122 Dialog.show();
123 zoomValue = Dialog.getNumber();
124 showInitialSelection = Dialog.getCheckbox();
125 surOri= Dialog.getNumber();
126 showDestination = Dialog.getCheckbox();
127 surZoom= Dialog.getNumber();
128 if (slices > 1) {
129     fromSlice= Dialog.getNumber(); FSlice=parseFloat (fromSlice);
130     toSlice= Dialog.getNumber(); TSlice=parseFloat (toSlice);
131 }
132 if (zoomValue < 1) zoomValue = 1;
133 if ((widthSel * zoomValue) >= width || (heightSel * zoomValue) >= height) {ok=0;} else {ok=1;
134 }
135 }
136
```

Paste
commands
here

Run

Kill

Show Errors

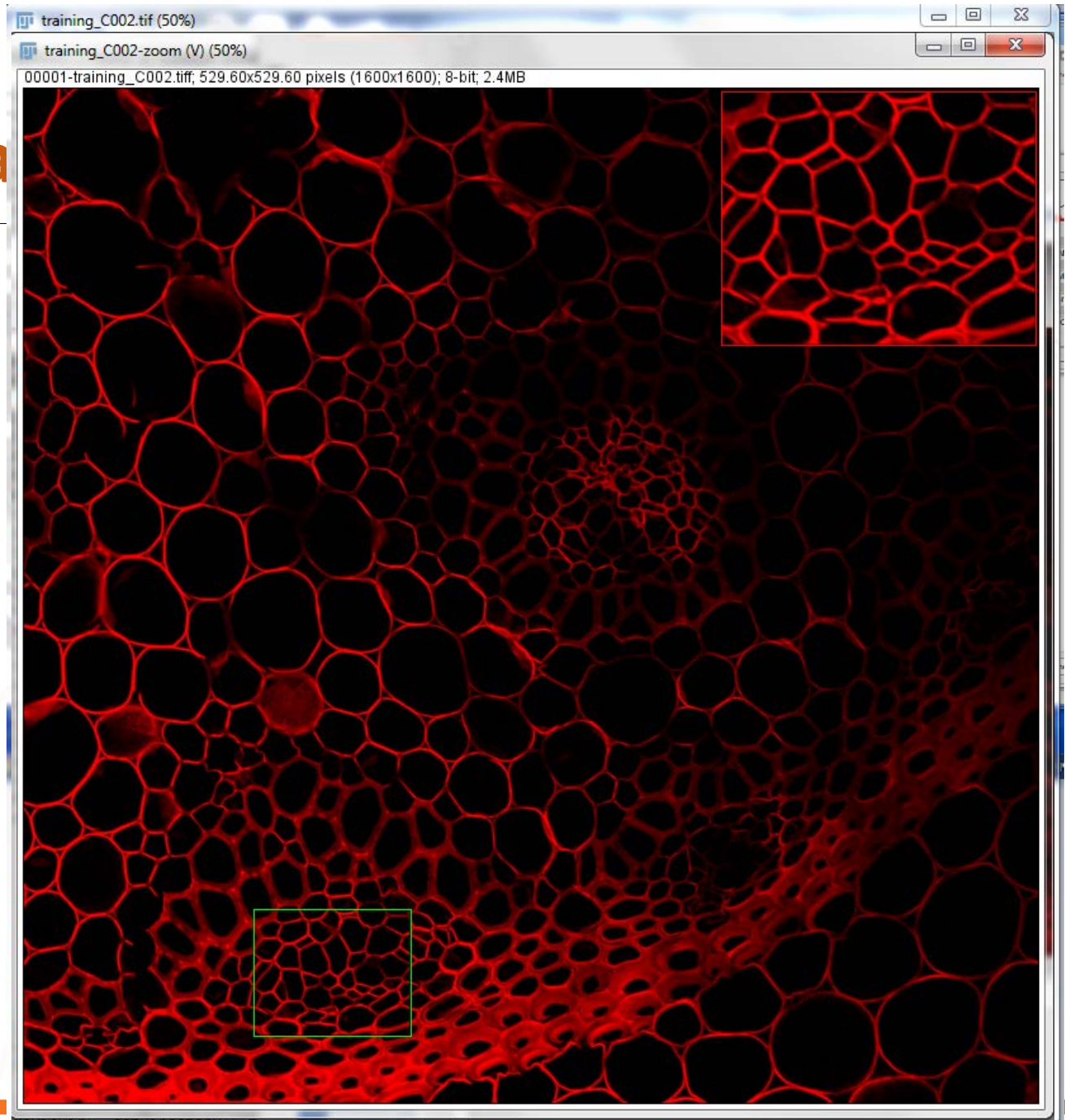
Clear

OK Cancel

Creating a

4. Now you decide where to place your inset (red square)

5. Finally click on the red square place the inset







Thank you!

Upcoming Imaging Events

When	What	Where
2:30-4:00 pm June 13	Westmead Imaging Community: New Products from Leica and Introduction to Cell Profiler	CMRI Seminar Room 1
11-5pm 14 June	User meeting: QC for acquisition & analysis Workshop on Olympus VS 120 image analysis	WIMR Conference Room Level 2
April-July ?	Art in Science	TBC
1-4.30pm 4 July	Seeing is believing	WIMR Conference Room Level 2
11-1pm 22 Nov?	Live cell imaging (TBC)	WIMR Conference Room Level 2