

Finding **cures**. Saving **lives**. Giving **hope**.



Colocalisation microscopy: basics, experiment setup and analysis

“Seeing is believing”

REALLY?

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Outlines

1. Defining Coloc

2. Influence on Coloc

3. Experiment setup: sample prep, acquisition, image processing

4. Evaluating Coloc

5. Data analysis: coefficients, available software

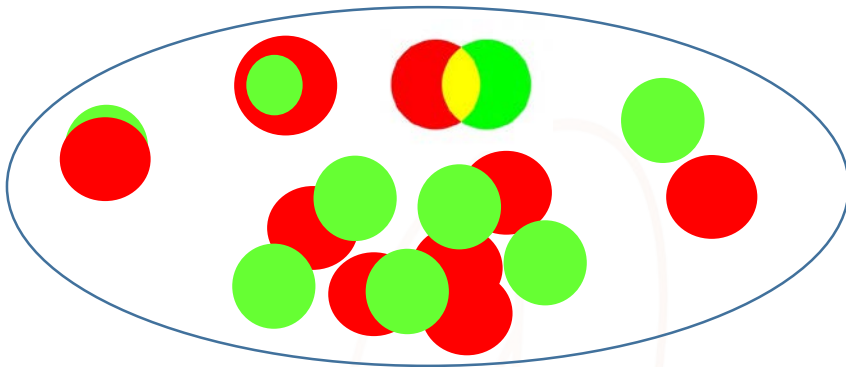
6. Coloc microscopy workflow

1. Defining colocalisation (Coloc)

Biology

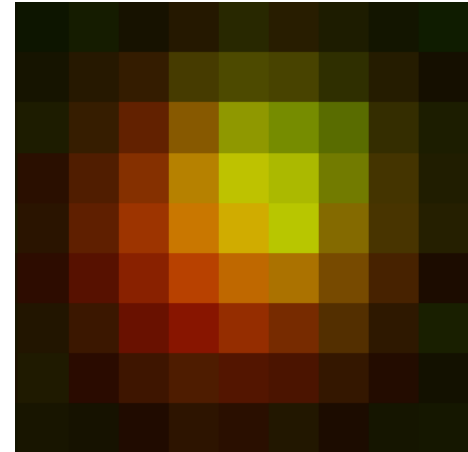
Co-occurrence (overlapping)

Correlation (closeness, association)



Digital imaging

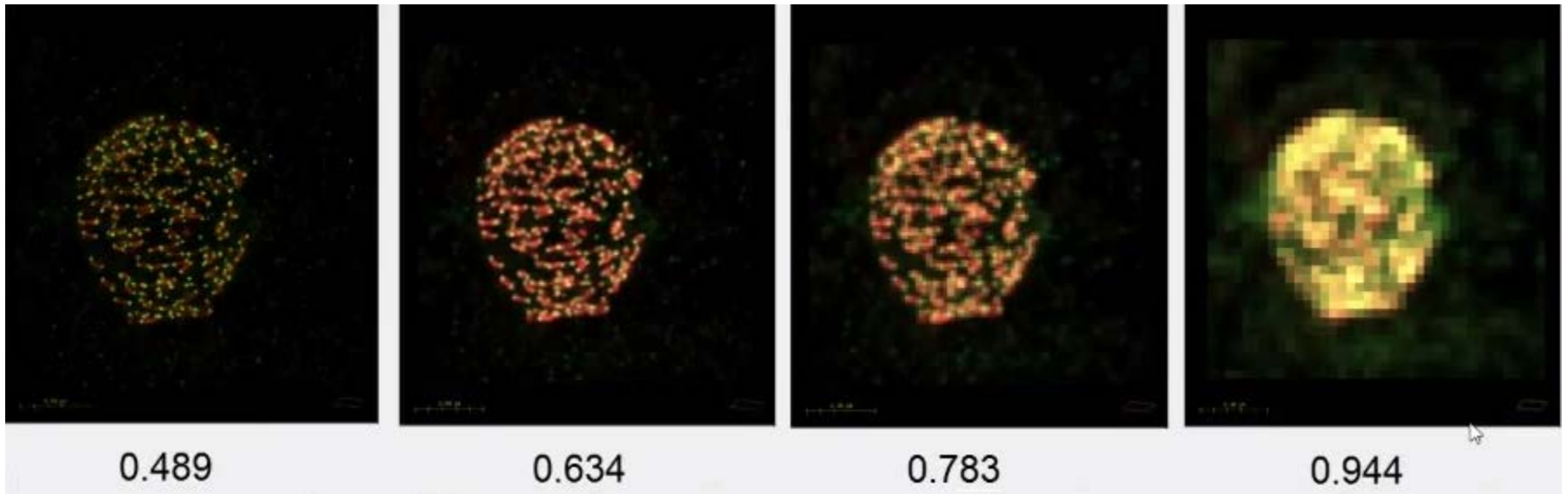
Sharing the same pixel location



- Coloc never measures interactions

2. Influence on Coloc—resolution

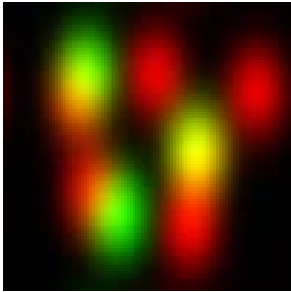
- Resolution



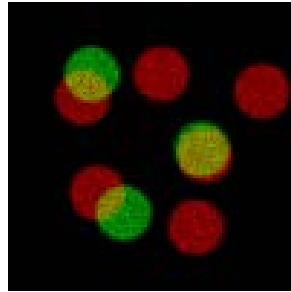
- High resolution Coloc microscopy is more convincing

2. Influence on Coloc—other difficulties

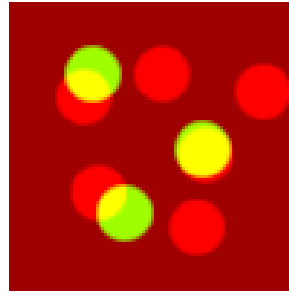
Resolution



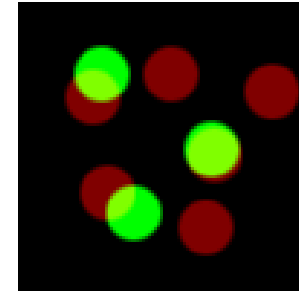
Noise



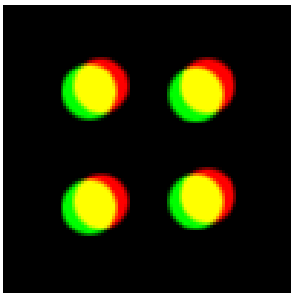
Background



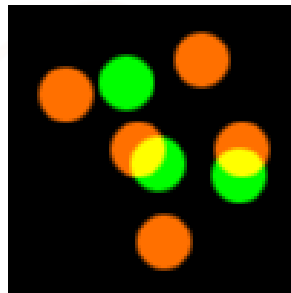
Intensity variations



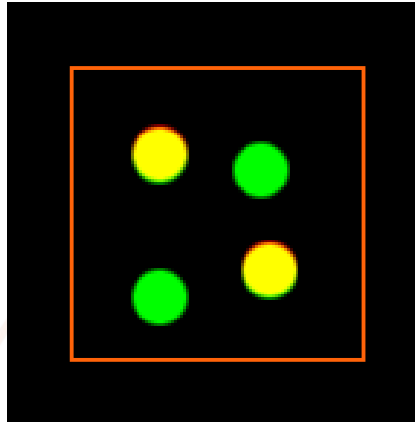
Cross-talk
Bleed-through



Chromatic
shift



Bkg margin



- Optimising experiment setup
- Image preparation helps: deconv, restoration etc

3. Experiment setup

Sample prep

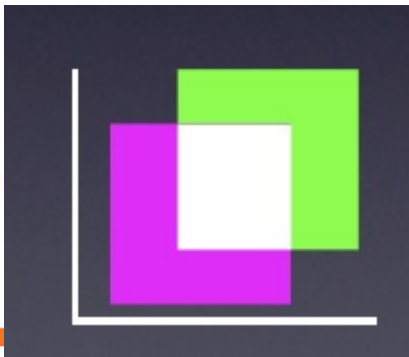
- Labelling optimisation
- AF
- Dye pair choice
- Controls

Mic & imaging

- Instrument choice
- Lens (NA, PlanApo, oil)
- Nyquist sampling
- **Z stacking**
- Low laser
- Seq, saturation
- Same conditions
- Beads imaging

Image processing

- Lossless format
- No change of Bitdepth
- No projection



Nyquist rate and PSF calculator

Microscope type	Confocal
Numerical aperture	1.3
Excitation wavelength	488 nm
Emission wavelength	520 nm
Number of excitation photons	1
Lens immersion refractive index	Oil 1.515

Calculate a Point Spread Function

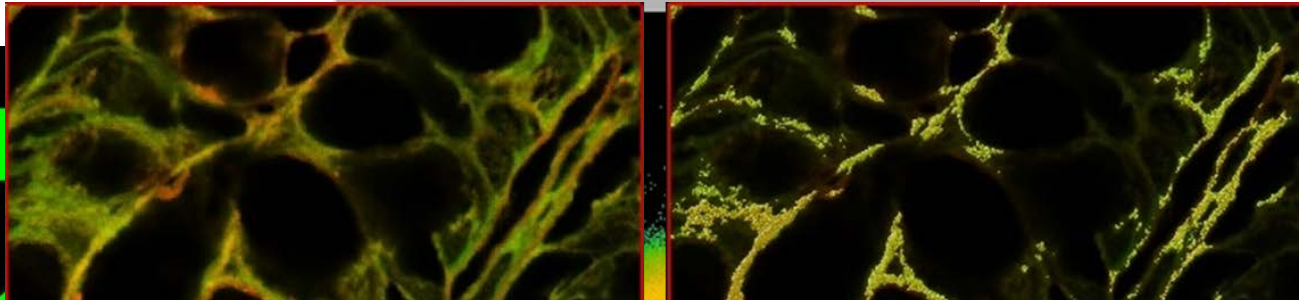
4. Evaluating Coloc

Qualitative/visual

- Color merge image
- Scatterplot
- Coloc map

Quantitative

- Object-based segmentation)
- Intensity-based (coefficients)



- Combine different methods
- Analyse multiple images + **statistics test**

5. Coloc analysis—image preparation

- **Image restorations (i.e. Huygens)**

cross talk

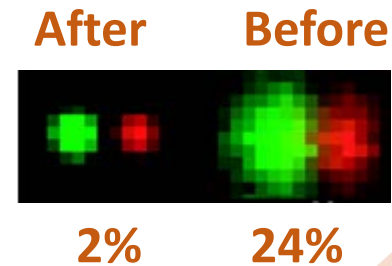
chromatic aberration

focal plane drift

others

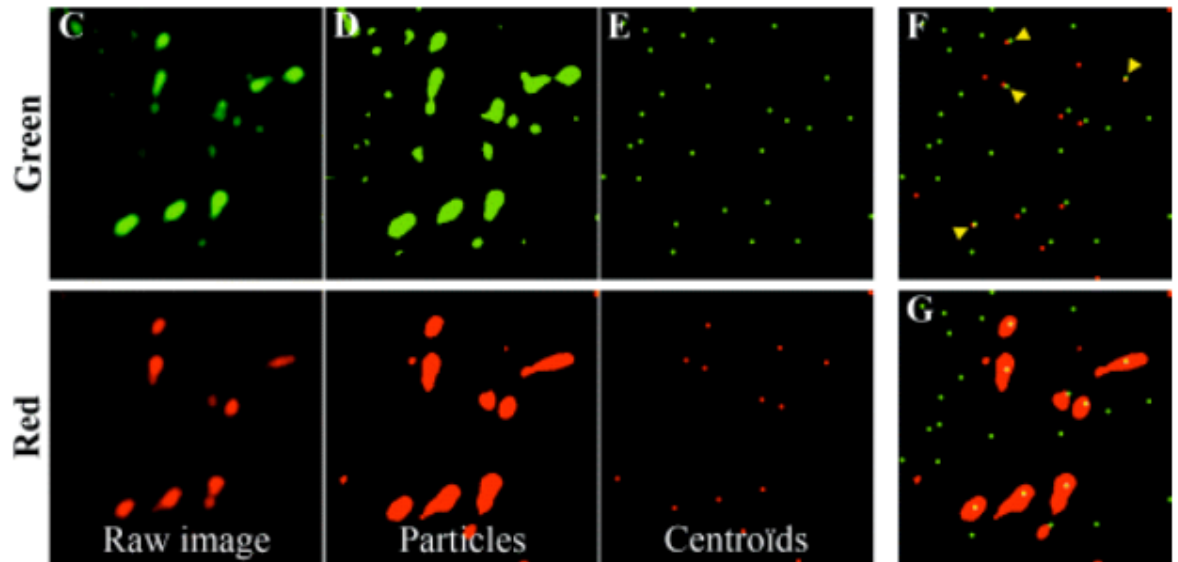
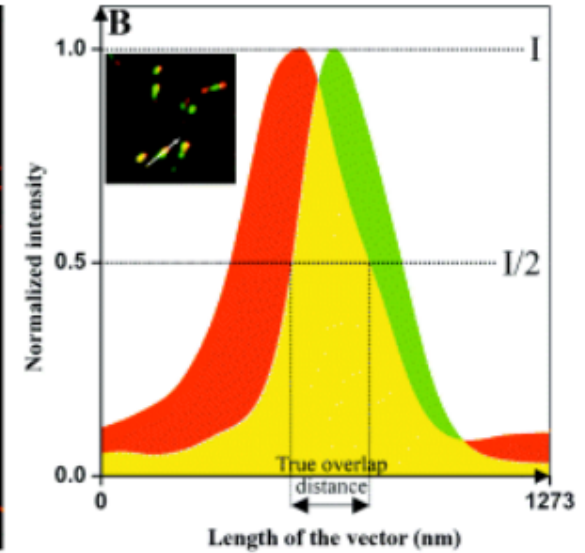
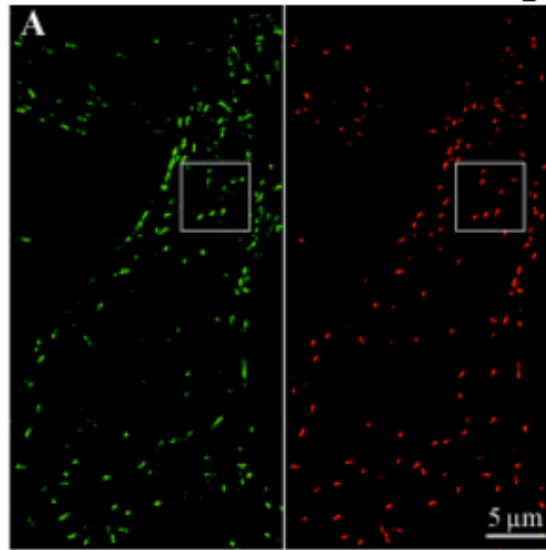
- **Deconvolution**

- **Background correction**



5. Coloc analy

- Large objects, se
- Less intensity de
- Measurements: c

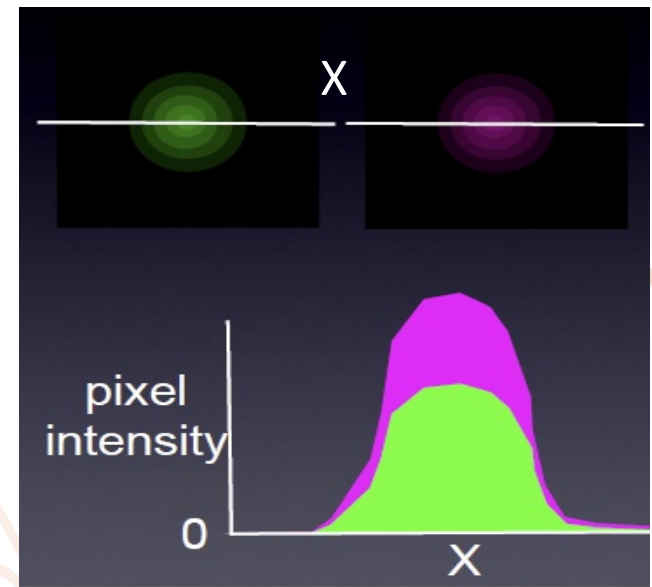


- Co-occurrence/ov

5. Coloc analysis---Intensity based analysis_1

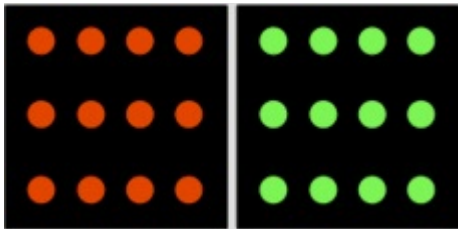
- Relationship between channel intensities
- Intensity profiles overlap
- Measurements

coefficients

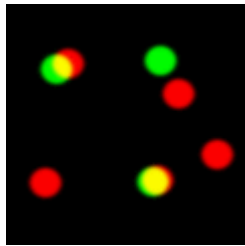


5. Coloc analysis---Intensity based analysis_2

100%

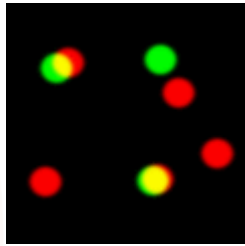
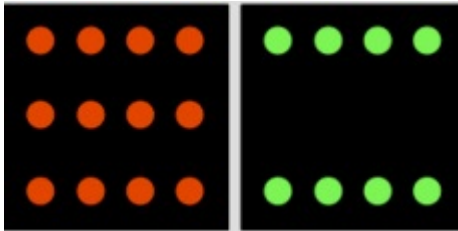


We need C are INSENSATIVE to...



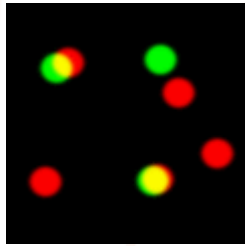
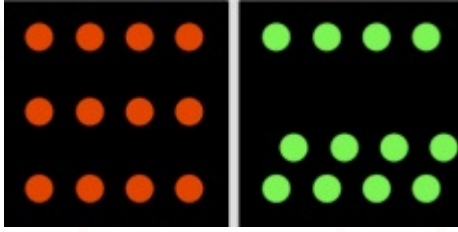
the addition of extra non-coloc

66% or 100?



relative variations of intensities

66%?



the presence of background

5. Coloc analysis---major coefficients for intensity based analysis

Which coefficient(s)? questions to ask...

- Correlation or co-occurrence?
- Random or true?

Correlation	Co-occurrence
1. Pearson's Correlation Coefficient (PCC)	1. Manders' Overlap Coefficients (MOC)
<ul style="list-style-type: none">• Object PCC	<ul style="list-style-type: none">• K_1, k_2
2. Spearman's Rank Correlation Coefficient (SRCC)	2. Manders' Coloc Coefficients
<ul style="list-style-type: none">• Object SRCC	<ul style="list-style-type: none">• $m_1, m_2; M_1, M_2$

Costes' Randomisation Analysis

5. Coloc analysis---Pearson's Correlation C_(PCC)

$$R_r = \frac{\sum_i (Ch1_i - Ch1_{mean}) \cdot (Ch2_i - Ch2_{mean})}{\sqrt{\sum_i (Ch1_i - Ch1_{mean})^2 \cdot \sum_i (Ch2_i - Ch2_{mean})^2}}$$

Pearson's Correlation Coefficient (PCC)

Meaning	the linear correlation of the intensity distribution between two channels. the similarity/correlation between shapes/pattern/variation of two channels, while ignoring signal intensities
Standard Values	-1~+1. 0: no correlation; >0: G is higher than Avg when R is higher than Avg (correlation); <0: G is lower than Avg when R is higher than Avg
Values indicating Colc	0.5-1
Values indicating absence of coloc	-1-0.5
Example	PC=0.8. 80% of pixels in both ch are positively correlated
Advantage	Not sensitive to the intensity of constant background Not sensitive to each channel/overlapping pixels (brightness)
Disadvantage	<ul style="list-style-type: none"> • Difficult to interpret • sensitive to non-coloc signals • No information about the individual channels. • Affected by noise
Application	Any coloc. Co-correlation. Sub-domains.

5. Coloc analysis---Spearman's Rank C (SRCC)

Spearman's Rank Correlation Coefficient (SRCC)

Meaning

SRCC is equivalent to the PCC, just to apply pixel intensity ranks (rank number 1, 2,3...) rather than to the intensities themselves

Standard Values

Values indicating Colc

Values indicating absence of coloc

Example

Advantage

None linear correlation

Disadvantage

Application

Non-linear correlation or codependance

5. Coloc analysis---Manders' Overlap C (MOC)

$R = \frac{\sum_i Ch1_i \cdot Ch2_i}{\sqrt{\sum_i (Ch1_i)^2 \cdot \sum_i (Ch2_i)^2}}$	<p>Overall Overlap Coefficient (R) also Manders' Overlap Coefficient (MOC)</p>
<p>Meaning</p>	<p>Indicates an actual overlap of the signals. the ratio of intersecting volume (ch1&2) to total object volume (Ch1&2)</p>
<p>Standard Values</p>	<p>0~+1. • R = 1 : perfect coloc/correlation • R= 0 : random (no) coloc</p>
<p>Values indicating Colc</p>	<p>>0.6</p>
<p>Values indicating absence of coloc</p>	<p><0.6</p>
<p>Example</p>	<p>R= 0.5 – 50% of both chan (objects) overlap</p>
<p>Advantage</p>	<p>Rough but easy to interpret • Not sensitive to the intensity of the overlapping pixels</p>
<p>Disadvantage</p>	<p>Sensitive to background & noise • No information about the individual ch</p>
<p>Application</p>	<p>Volume in each ch is more or less the same and intensities are constant inside the objects and the image can be considered as binary (B/W).</p>

5. Coloc analysis---Manders' K overlap C (k1, k2)

$$k_1 = \frac{\sum_i Ch1_i \cdot Ch2_i}{\sum_i (Ch1_i)^2} \quad k_2 = \frac{\sum_i Ch1_i \cdot Ch2_i}{\sum_i (Ch2_i)^2}$$

$$R^2 = k_1 \cdot k_2$$

The k overlap coefficients (k1, k2)

Meaning	R2=k1• k2 . Split the value of coloc into two separate parameters, allows to determine the contribution of each ch/Ag to the areas with coloc
Standard Values	vary
Values indicating Colc	>0.5
Values indicating absence of coloc	Close values (0.5 and 0.6; 0.8 and 0.9)
Example	Distant values (0.5 and 0.9; 0.2 and 0.7)
Advantage	The 2 channels are analyzed separately • Addition of a not colocalized signal will affect only one of the channels
Disadvantage	depend strongly on the ratio of total intensities in both channels
Application	When two ch intensities are at a similar level

5. Coloc analysis---Manders' Coloc C (m1, m2)

$$m_1 = \frac{\sum_i Ch1_{i,coloc}}{\sum_i Ch1_i} \quad m_2 = \frac{\sum_i Ch2_{i,coloc}}{\sum_i Ch2_i}$$

Manders' original coefficients (m1, m2)

Meaning	Describes portion of each ch coloc with the other. % of Ch1 pix intensities coloc with Ch2 (m1).
Standard Values	0~+1.
Values indicating Colc	0.5-1
Values indicating absence of coloc	<0.5
Example	m1=1, m2=0.4: 100% of Ch1 pixel intensities coloc/overlap with Ch2, but only 40% of Ch2 pixel intensities coloc/overlap with Ch1.
Advantage	independent of signal proportionality (good for non linear); provides two components
Disadvantage	Sensitive to background
Application	Any coloc.

5. Coloc analysis---Manders' Coloc C (M1, M2)

$$M_1 = \frac{\sum_i Ch1_{i,coloc}}{\sum_i Ch1_i} \quad M_2 = \frac{\sum_i Ch2_{i,coloc}}{\sum_i Ch2_i}$$

Manders' Thresholded Coefficients (M1, M2)

Meaning	% of Ch1 pix intensities coloc with Ch2 (m1). Same as m1 and m2 but applied to analyze scatter gram ROI. Thus, M1 can be defined as the co-occurrence fraction of color 1 with color 2. Likewise, M2 is the co-occurrence fraction of color 2 with color 1.
Standard Values	0~+1.
Values indicating Colc	0.5-1
Values indicating absence of coloc	<0.5
Example	M1=1, M2=0.4: 100% of Ch1 pixel intensities coloc/overlap with Ch2, but only 40% of Ch2 pixel intensities coloc/overlap with Ch1.
Advantage	Less sensitive to background problems. less dependent on the actual intensity ratios between channels
Disadvantage	Difficult to interpret • No information about the individual channels • Affected by noise
Application	Any coloc.

Statistics by Region Thresh. Thresh.

ROI	Area (pixels^2)	Pearson's Coeff.	Overlap	Overlap Index 1	Overlap Index 2	Coloc. Index 1	Coloc. Index 2
Entire Series	2.56e+006	0.92719	0.95875	0.78612	1.1693	0.34168	0.6197

Mode

Objects

Colocalization <3>

Input Output

Select Region

Input Channels Thres

475/523

542/594

Pearson Coefficient of Correlation

Colocalization - gut .oif Z:0 T:0 L:0

Data Selection

Single Series

Axis

Vertical Horizontal

AnnotationMode

	Threshold
V	296
H	231

ExtractArea

Annotation		
Lower-left(C)	45.62 [%]	ofAll Samples
	159124 [pixel]	samples
Lower-right(D)	11.60 [%]	Right<VerticalThreshold
	11.99 [%]	Lower>=HorizontalThreshold
	6.22 [%]	ofAll Samples

Twist 360

Select statistics:

#	Label	Chan	F
3	1	1	
4	2	1	
5	3	1	

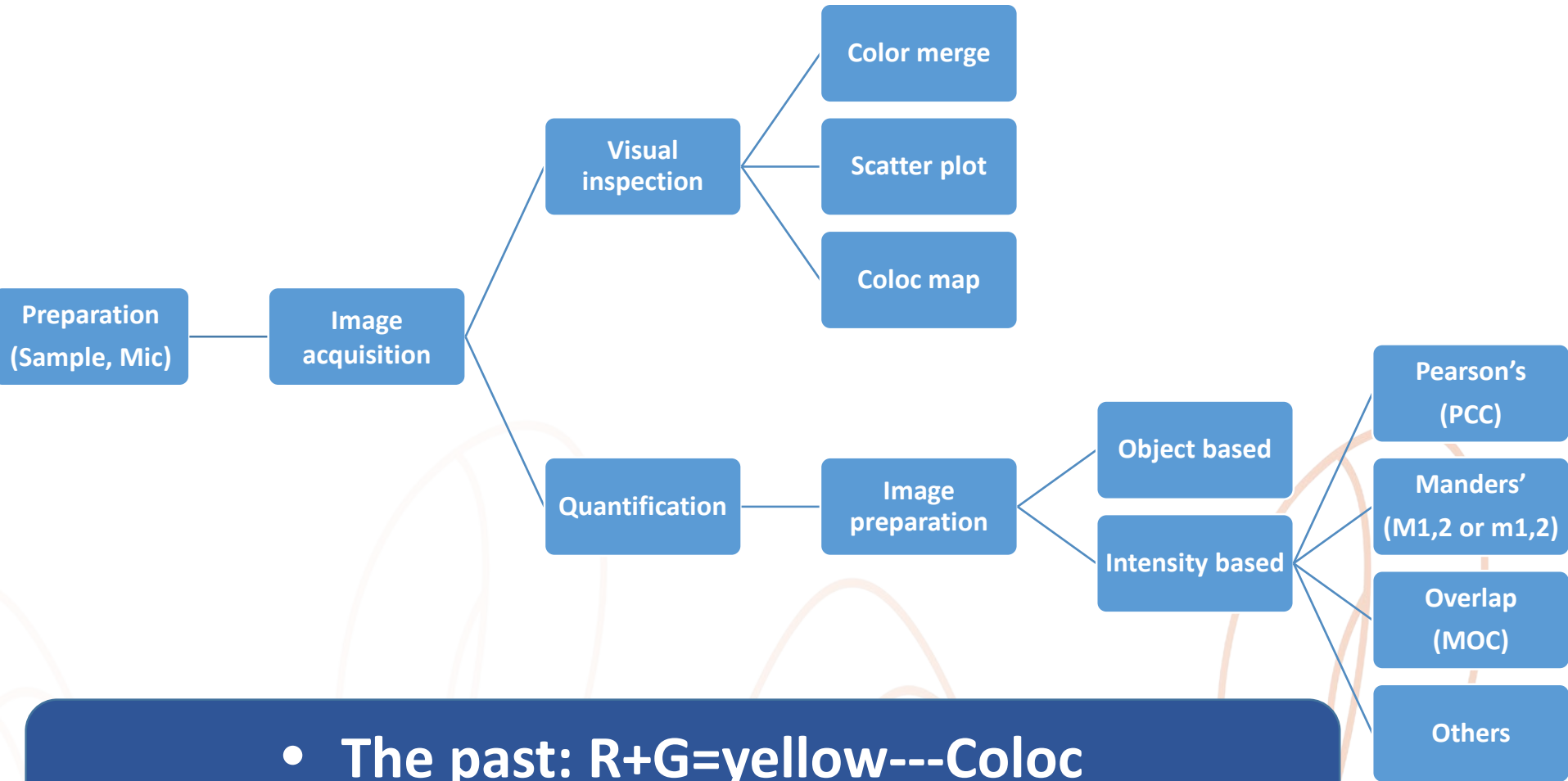
Info

1

2

3

6. Coloc microscopy workflow



- The past: R+G=yellow---Coloc
- From now on: 3D, Decon, def, quantif, stat...



Acknowledgement & References

<https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.1365-2818.2006.01706.x>

<https://svi.nl/ColocalizationCoefficients>

<http://jcs.biologists.org/content/joces/131/3/jcs211847.full.pdf>

[And many others...](#)

Thank you!